

Physical and mechanical properties of sago starch – alginate films incorporated with calcium chloride

*Fazilah, A., Maizura, M., Abd Karim, A., Bhupinder, K., Rajeev Bhat, Uthumporn, U. and Chew, S. H.

Food Technology Division, School of Industrial Technology, University Sains Malaysia, 11800 Penang, Malaysia

Abstract: Edible films were prepared from sago starch: alginate mixtures with ratios of 1:0, 4:1, 3:1, 2:1, 1:1 and 0:1. The physical and mechanical properties of films were modified with the addition of calcium chloride (CaCl_2) (2×10^4 mg/kg) in the film solutions and by immersion of the films into a CaCl_2 (2% w/w) solution. The films were characterized for tensile strength (TS), percentage of elongation at break (% E), calcium content and water solubility (WS). The calcium content in 100% alginate films was significantly higher compared to those after immersion for 1 min in a CaCl_2 solution. Immersion of films into a CaCl_2 solution for 1 min increased the TS, except for 100% sago starch film. Prolonging the immersion time for 10 minutes did not significantly affect the TS of the films. Moreover, increasing the immersion time did not significantly decrease the WS of the films. Addition of CaCl_2 in film solutions with higher ratios of alginate significantly increased their TS and % E. Films treated with CaCl_2 by the immersion process led to markedly increased TS compared to those treated by CaCl_2 addition in the film solutions.

Keywords: Alginate, calcium chloride, mechanical properties, physical properties, sago starch

Introduction

The production of edible films from natural polymers has received much attention due to the excellent biodegradability, biocompatibility and edibility of the films. Preparing these films involves the use of at least one film-forming agent (macromolecules), a solvent and a plasticizer. The most-used macromolecules are polysaccharides, proteins and lipids (Krochta and Mulder-Johnston, 1997; Cuq *et al.*, 1998; Krochta, 2002). There are many polysaccharide sources for the production of edible films, such as root and cereal starches, plant cell pectin, alginate and carrageenan from seaweed and cellulose (Kester and Fennema, 1986). However, starches are commonly used as a raw material because they are renewable resources, widely available and inexpensive (Mali and Grossmann, 2003; Famá *et al.*, 2005).

Starch is a major food component and a complex biodegradable carbohydrate made up of thousands of glucose units (Zhou *et al.*, 2009). It consists primarily of linear and branched chains of glucose molecules, namely amylose and amylopectin. Amylose, which is the linear fraction of starch, is responsible for the formation of coherent, relatively strong and free-standing films (Rindlav-Westling *et al.*, 1998). Physical crosslinkages in the macromolecular network of starch are formed mainly by microcrystalline amylose and affect the mechanical properties of films (Rindlav-Westling *et al.*, 1998). In contrast, the

branched structure amylopectin generally leads to films that are brittle and reduced tensile strength.

Alginate is the water-soluble salt of alginic acid. It is a naturally occurring, non-toxic polysaccharide found in brown algae (Rubio and Ghaly, 1994; Al-Musa *et al.*, 1999) and consists of β -(1-4)-linked D-mannuronic acid (M) and α -(1-4) linked L-guluronic acid (G). Three types of glycosidic linkages exist in the block structures: diequatorial M-M, diaxial G-G and dequatorial-axial MG (Yotsuyanagi *et al.*, 1991; Donati *et al.*, 2005; Dong *et al.*, 2006). Alginate has potential use in films or as a coating component because of its unique colloidal and gel-forming properties (King, 1982; Rhim, 2004). However, the most useful and unique property of alginate is its ability to react with polyvalent metal cations, such as calcium ions in aqueous media, to generate strong gels and insoluble polymers (Grant *et al.*, 1973; Braccini and Pérez, 2001; Fang *et al.*, 2008; Olivas and Barbosa-Cánovas, 2008). As such, several studies have analyzed the properties of alginate-calcium films (Pavlath *et al.*, 1999; Rhim, 2004; Olivas and Barbosa-Cánovas, 2008; Silva *et al.*, 2009).

Starch-based films exhibit physical characteristics similar to synthetic polymers, such as being transparent (Lourdin *et al.*, 1997; Myllärinen *et al.*, 2002; Parra *et al.*, 2004), odorless, tasteless (Wolff *et al.*, 1951), semi-permeable to carbon dioxide and resistant to oxygen passage (Nisperos-Carriedo, 1994). In addition, they have good barrier and mechanical

*Corresponding author.
Email: fazilah@usm.my
Fax: +604 6573678

properties (Myllärinen *et al.*, 2002; Larotonda *et al.*, 2005). Several researchers have tried to develop more sophisticated starch-based edible films with enhanced barrier and mechanical properties by blending them with other biopolymers, such as alginate (Wu *et al.*, 2001), potato starch with agar (Wu *et al.*, 2009), cassava starch with arabinoxylan (Phan The *et al.*, 2009), and tapioca starch with hsian-tsaio leaf gum (Chen and Lai, 2008).

Wu *et al.* (2001) and Maizura *et al.* (2007) studied on starch-alginate mixtures to form edible films. However, sago starch-alginate mixtures in the presence of CaCl_2 have not been reported for use in the formation of edible films. Starch-alginate mixtures crosslinked with calcium ions are desirable to produce films with enhanced tensile strength and decreased solubility. The objective of this research was to investigate the effect of CaCl_2 on the physical and mechanical properties of sago starch-alginate edible films.

Materials and Methods

Sago starch (*Metroxylon sagu*) was obtained from Nitsei Sago Industries, Sdn. Bhd. Penang, Malaysia. Alginate (Acros Organic, Fairlawn, N.J., U.S.A.), glycerol (Qrec, New Zealand) and calcium chloride (System, Shah Alam, Selangor, Malaysia) were also used in this study.

Preparation of sago starch-alginate film

Film-forming solutions were prepared from mixtures of sago starch and sodium alginate containing different ratios (1:0, 4:1, 3:1, 2:1, 1:1, 0:1) based on the total mass of dry solid (3.5 g) and glycerol (0.5 g) in 200 ml of distilled water. Films were treated with CaCl_2 in two different ways. In the first treatment, 2×10^4 mg/kg of CaCl_2 was mixed into the film solutions. The mixture was heated to 85°C with continuous stirring for 45 min, followed by homogenization at 1000 $1/s$ using a homogenizer (IKA-Werke GmbH, Staufen, Germany) for 5 min. The solution (90 g) was cast onto a 16 x 16 cm polyacrylic plate, cooled down to room temperature, and oven-dried at 40°C for 24 hr. For the second treatment, sago starch-alginate films were immersed in a 2 % w/w CaCl_2 solution for 1, 5 and 10 min and dried at 40°C for 6 hr.

Film thickness

The film thickness was measured using a micrometer (Dial thickness gauge 7301, Mitutoyo Co. Tokyo, Japan) with accuracy to the nearest 0.01 mm at five locations along the film strips; the average value was used in tensile strength calculations.

Mechanical properties

The tensile strength and elongation at break percentage of films were tested using a TA-TX2 texture analyzer (Stable Micro System, Goldaming, Surrey, UK) in accordance with ASTM (ASTM, 1981). Five sample strips (14.0 x 2.0 cm) of each formulation were cut and clamped between tensile grips. Films were previously equilibrated in 52% relative humidity (RH) at 30°C for 7 days. Each film strip was placed in pneumatic grips (30 psi) and stretched at 1.5 mm s^{-1} . The initial distance between the grips was 100 mm. The tensile stress (Mpa) and percentage of elongation at break (%E) were determined. The tensile strength of the films is a measure of the maximal force per original cross-sectional area that the film could sustain before breaking, and the elongation at break percentage measures the capacity of the film to extend before breaking (Silva *et al.*, 2009). At least five strips were used per treatment, with five replicates.

Calcium content

The concentration of calcium in the crosslinked films was determined with an atomic absorption spectrophotometer (Perkin Elmer, Waltham, Massachusetts, U.S.A.). The film samples (1 g) were dissolved in 6 mL of a concentrated nitric acid (HNO_3) solution and 1 mL of 30% (v/v) hydrogen peroxide (H_2O_2). The mixture was digested using a microwave digester for 20 min. The clear solutions were then mixed with 5 mL of lanthanum oxide (La_2O_3), and distilled deionized water was added until the solution volume was 25 mL. Each determination was done in triplicate.

Staining microscopy

Dried film samples (1.5 x 2 cm) were placed onto a glass slide. Three drops of an iodine solution were added on the films. The films were left for a few seconds to allow the iodine solution to completely disperse on the films before covering with a cover slip. Then, the film structure was examined using light microscopy.

Water solubility

The water solubility of the films was determined according to the method described by Romero-Bestida *et al.* (Romero-Bastida *et al.*, 2005). Pieces of film (2 x 3 cm) were cut from each film formulation and stored in a dessicator with silica gel (0% RH) for 7 days. The samples were weighed to the nearest 0.0001 g and placed in beakers with 80 ml of deionized water. The samples were maintained under constant agitation for 1 hr at room temperature (approximately 25°C). After

soaking, the remaining pieces of film were filtered through filter paper (Whatman No. 1), followed by oven drying at 60°C to constant weight. The samples were measured in triplicate and the percentage of total soluble matter (% solubility) was calculated according to Eq. (1):

$$\% \text{ solubility} = \frac{(\text{initial dry weight} - \text{final dry weight}) \times 100}{\text{initial dry weight}} \quad (1)$$

Statistical analysis

The mean data was analyzed by two-way and one-way analysis of variance (ANOVA) to evaluate the effects of CaCl₂ and the starch alginate ratios on the physical and mechanical properties of the films using the SPSS 15.0 system software. Duncan's multiple-range test (P<0.05) was used to determine the significant differences between means.

Results and Discussions

Whereas the starch films appeared translucent, stiff, slightly brittle, odorless and colorless, the alginate films were transparent and pliable. Mixtures of these two polymers formed films that were almost transparent, had good flexibility and were easily removed from the casting plate. The viscosity of the film-forming solution with higher amounts of alginate increased after the addition of CaCl₂, likely due to the development of crosslinks between the carboxyl groups in the film molecules and the calcium ions. After the immersion of films into the CaCl₂ solution, the surface was not as smooth as that of the control films. A similar occurrence was also observed by Rhim (2004). These results were attributed to disruption of the film matrix during the immersion and an uneven rate of water removal from the films during the drying process.

The film thickness with different treatments of CaCl₂ and different starch alginate ratios is presented in Table 1. The thickness of films with sago starch: alginate ratios of 1:0 and 4:1 decreased significantly (P<0.05) after immersion in the CaCl₂ solution for

5 min and 10 min respectively compared to films without treatment with CaCl₂. These results suggest that the starch component in the starch-dominated films solubilized into the solution, thus reducing the thickness of the film. However, no significant difference (P>0.05) in thickness was observed in films with higher ratios of alginate (3:1, 2:1, 1:1 and 0:1) after immersion in a CaCl₂ solution for 10 min compared to films without CaCl₂ (control).

The mechanical properties of edible films are of great importance due to their influence on product performance. The effect of CaCl₂ on the tensile strength of the films at different sago starch: alginate ratios are shown in Figure 1. The results show that films without the addition of alginate had lower tensile strengths compared to those with alginate. Many studies (Romero-Bastida *et al.*, 2005; Turhan and Sahbaz 2004; Laohakunjit and Noomhorm 2004; Fama *et al.*, 2005; Mali *et al.*, 2005) was done on the tensile strength analysis of starch based films. The tensile strength of sago starch based films obtained in this study was similar as found by Romero-Bastida *et al.* (2005) for banana starch film which was 25 Mpa. However, the presence of alginate at sago starch: alginate ratios 4:1 had significantly increased the tensile strength due to the linear structure of the polymer, which is capable of producing tough and flexible films. In addition, films with sago starch: alginate ratios 2:1, 1:1 and 0:1 exhibited increased tensile strength by more than 40% compared to films without alginate.

The results in Figure 1 also clearly indicate that the tensile strength of films with CaCl₂ significantly increased (P<0.05) compared to those films without CaCl₂ at sago starch: alginate ratios of 2:1, 1:1 and 0:1. These results could be attributed to crosslinking between carboxyl groups in alginate with calcium ions. According to Glickmans (1982), crosslinks could be formed by simple ionic bridging of two carboxyl groups on adjacent polymer chains with calcium ions. However, calcium ions could also form connections with either polymannuronic acid or polyguluronic acid segments.

Table 1. Effect of calcium chloride and starch:alginate ratios on film thickness

Treatment	Thickness (mm) x 10 ⁻²					
	Sago starch: alginate ratio					
	(1 : 0)	(4 : 1)	(3 : 1)	(2 : 1)	(1 : 1)	(0 : 1)
Control (films without CaCl ₂)	12.8 ± 0.4 ^{bijk}	12.4 ± 0.5 ^{fg hij}	11.6 ± 0.5 ^{cdefg}	12.0 ± 0.7 ^{defghi}	11.4 ± 0.5 ^{cdef}	13.4 ± 0.5 ^{ijkl}
Films incorporated with CaCl ₂	12.4 ± 0.5 ^{fg hij}	12.6 ± 0.5 ^{ghij}	12.4 ± 0.5 ^{fg hij}	12.2 ± 0.4 ^{efghi}	11.6 ± 0.5 ^{cdefg}	12.6 ± 0.5 ^{ghij}
Films immersed in CaCl ₂						
1 min	11.8 ± 0.4 ^{defgh}	11.6 ± 0.9 ^{cdefg}	11.4 ± 0.9 ^{cdef}	10.6 ± 0.9 ^{abc}	11.2 ± 0.8 ^{bcde}	13.0 ± 1.4 ^{ijkl}
5 min	11.4 ± 1.3 ^{cdef}	11.4 ± 0.5 ^{cdef}	11.0 ± 1.0 ^{bed}	11.4 ± 0.9 ^{cdef}	11.0 ± 1.2 ^{bed}	14.0 ± 0.7 ⁱ
10 min	9.8 ± 0.4 ^a	10.2 ± 0.4 ^{ab}	11.8 ± 0.4 ^{defgh}	11.8 ± 0.4 ^{defgh}	12.2 ± 1.1 ^{efghi}	13.8 ± 0.4 ^{kl}

Mean ± standard deviation (n= 5); different superscript letters in each row and column indicate significant differences at (P < 0.05).

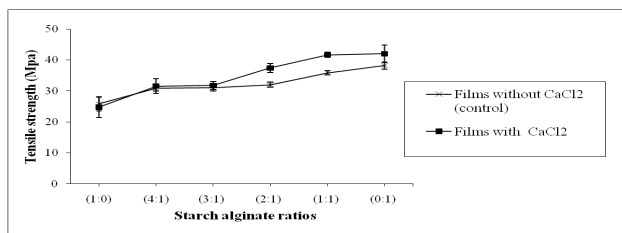


Figure 1. Effect of starch:alginate ratios on tensile strength of films with and without CaCl₂. The vertical bars are standard deviations of the means (n=5).

The tensile strength of films with various ratios of starch and alginate after immersion in the CaCl₂ solution for different periods of time is presented in Figure 2. The tensile strength of the films drastically increased after immersion for 1 min in the CaCl₂ solution. After immersion in the CaCl₂ solution for 1 min, films with a sago starch: alginate ratio of 4:1 had higher tensile strengths (39.9 ± 3.0 Mpa) compared to films (30.8 ± 0.8 Mpa) with similar sago starch: alginate ratios incorporated with CaCl₂. These results suggest that the effect of increased tensile strength is more pronounced in the immersion process. The calcium ions present in the solution can crosslink with alginate in the films matrix better than calcium ions and alginate in the edible film solutions at similar concentrations of 2% w/w CaCl₂. However, there was no significant (P>0.05) increase in the film tensile strength after immersion in the CaCl₂ solution for 10 min.

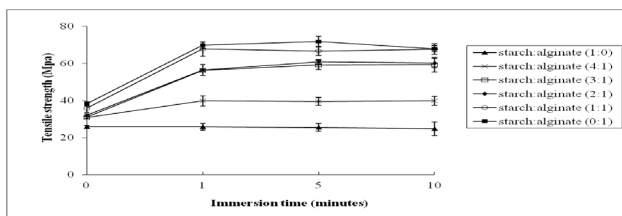


Figure 2. Effect of starch:alginate ratios on tensile strength of films after immersion in CaCl₂ for different periods of time.

The effects of CaCl₂ on the percentage of elongation at break of the films at different sago starch: alginate ratios are shown in Figure 3. The elongation at break sharply increased in films with a 0:1 ratio (sago starch: alginate), both with and without CaCl₂. Generally, increased tensile strength led to a decreased percentage of elongation at break (Rhim, 2004; Olivas and Barbosa-Cánovas, 2008). However, we observed that increased tensile strength of films did not necessarily decrease the percentage of elongation at break. These results are similar to those obtained by Pavlath *et al.* (1999).

Figure 4 shows the percentage of elongation at break of films with various ratios of sago starch and alginate after immersion in CaCl₂ solutions for different periods of time. Longer immersion times

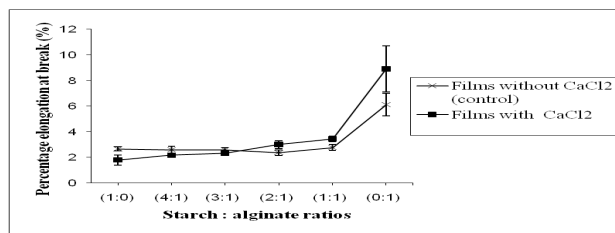


Figure 3. Effect of starch:alginate ratios on percentage of elongation at break of films with and without CaCl₂. The vertical bars are the standard deviations of the means (n=5).

did not significantly (P>0.05) increase the elongation at break percentage of all the films. The elongation of alginate films was substantially higher compared to other films. A study by Rhim (2004) found that, the elongation at break for alginate films was 14.0% and after immersion in 2% w/w of CaCl₂ for 5 mins, the percentage of elongation had decreased significantly to 3.4%. In this study, it was similarly found that immersion of alginate film for 5 mins also had decreased the elongation at break significantly from 8.90% to 7.46%. The films dominated with alginate exhibited higher tensile strength and good flexibility.

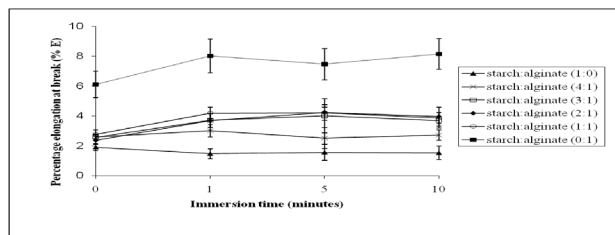


Figure 4. Effect of starch:alginate ratios on percentage of elongation at break of films after immersion in CaCl₂ for different periods of time. The vertical bars are the standard deviations of the means (n=5).

Staining microscopy analysis was conducted to determine the homogeneity of the produced films. Figure 5 shows films with different treatment of CaCl₂. The mixture of sago starch and alginate produced films with a smooth surface. However, incorporation of CaCl₂ into the sago starch and alginate film solutions led to the formation of films that were slightly rough and had a gritty surface, which could be related to the rapid rate of alginate gelation. In the absence of a sequestering agent for calcium, the gelation of the alginate component in a starch-alginate mixture would occur very fast (uncontrolled), thus preventing uniform formation of the film. In the immersion process, calcium ions crosslinked with alginate in the film matrix. We assumed that the presence of calcium ions in the film matrix did not interrupt the sago starch-alginate polymer chain crosslinking, thereby producing films with slightly smooth surfaces. After immersion in the CaCl₂ solution, these films dried much faster than the control films, and a similar finding was also reported

by Rhim (2004). However, the obtained films were unable to revert back to their smooth original surfaces after the immersion process, which may be due to the variable drying process between different parts of the film surface.

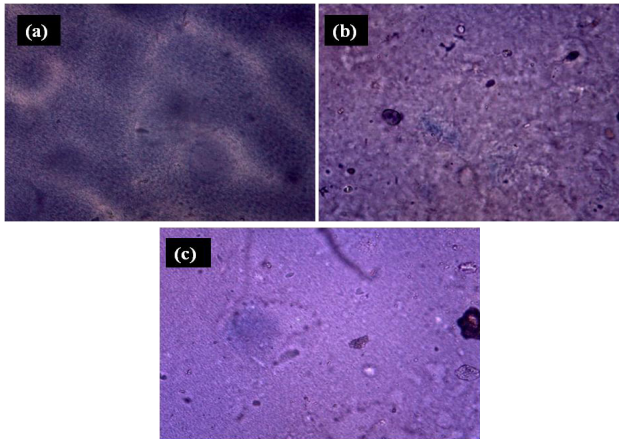


Figure 5. Representative pictures of sago starch-alginate (1:1) films with and without CaCl_2 treatment: (a) Control (without CaCl_2), (b) Film with addition of 2% w/w CaCl_2 , (c) Film immersed in 2% w/w CaCl_2 for 1 min.

The calcium content of films with different sago starch: alginate ratios after immersion in CaCl_2 solution for different periods of time is presented in Figure 6. Increasing the immersion time did not significantly ($P>0.05$) increase the calcium content in the films. The calcium ions probably crosslinked with almost all of the alginate polymers in the film matrix after 1 min of immersion time. Films with higher ratios of alginate have been predicted to have higher calcium content because of the mechanical properties of films (Olivas and Barbosa-Cánovas, 2008). Films with higher calcium content have more crosslinkages between calcium ions and alginate polymers, thus producing films with higher tensile strength. This relationship is supported by the significantly positive correlation between the tensile strength and calcium content values of films after 1 min of immersion in the CaCl_2 solution. The correlation value was 0.681, which is significant at the $P<0.01$ level.

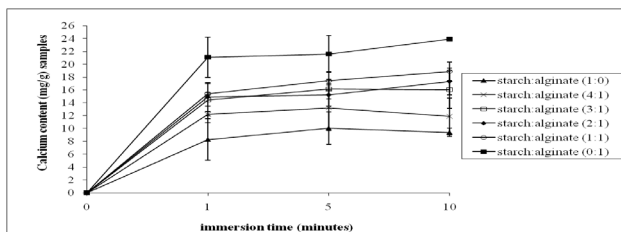


Figure 6. Effect of starch:alginate ratios on calcium content of films after immersion in a CaCl_2 solution for different periods of time. The vertical bars are the standard deviations of the means ($n=3$).

Water solubility is an important property of edible films. Some applications may require water

insolubility to enhance product integrity and water resistance. The results of the percentage of water solubility of films at different ratios of sago starch and alginate with and without CaCl_2 are shown in Figure 7. Films with CaCl_2 had higher water solubility compared to those without CaCl_2 at all ratios. The absence of a sequestering agent for calcium produced non-uniform films due to the rapid rate of alginate gelation, thus producing non-homogeneous film structures that disintegrated rapidly during immersion in water. In addition, films with CaCl_2 exhibited sharply increased water solubility as the alginate ratio increased. Alginate films with and without CaCl_2 completely disintegrated in water, making it impossible to measure the water solubility, as similarly noted by Rhim (2004).

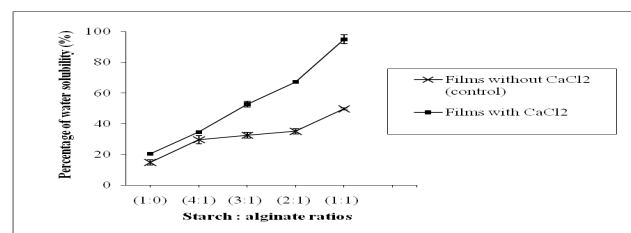


Figure 7. Effect of starch:alginate ratios on water solubility of films with and without CaCl_2 . The vertical bars are the standard deviations of the means ($n=3$).

The water solubility of films with various sago starch:alginate ratios at different periods of immersion times is presented in Figure 8. Increasing the immersion time in CaCl_2 solution did not significantly ($P>0.05$) enhance the film solubility in water. According to Pavlath *et al.* (1999), alginate films were not moisture-resistant and readily dissolved in distilled water within a minute. However, increases in water resistance of the films are dependent on the immersion time and concentration of calcium chloride (Pavlath *et al.*, 1999, Rhim, 2004). In this study, 100% alginate film, after immersion for 1, 5 and 10 min in a CaCl_2 solution (2%w/w), dissolved completely in water. Our results clearly showed that films with higher ratios of sago starch exhibited lower water solubility, indicating higher water resistance.

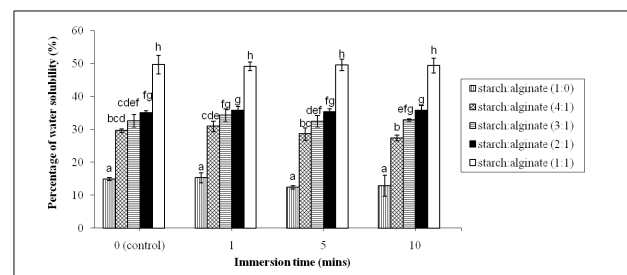


Figure 8. Water solubility of films with various starch:alginate ratios for different immersion times. The vertical bars are the standard deviations of the means ($n=3$). Different superscript letters within and across the group are significantly different at $P<0.05$.

In conclusion, our results demonstrated that mixtures of sago starch and alginate produced films with better tensile strength compared to sago starch films. However, immersion of sago starch-alginate films in CaCl_2 solution exhibited higher tensile strength than those with added CaCl_2 . After immersion in CaCl_2 solutions, films with different ratios of sago starch and alginate had better flexibility sago starch film. The films with higher ratios of alginate had higher calcium contents. Alginate film had a lower water resistance compared to sago starch film. Sago starch-alginate films had moderate water solubility.

Acknowledgment

This work was supported by a short-term grant (304/PTEKIND/636048) from University Sains Malaysia.

References

- Al-Musa, S., Abu Fara, D. and Badwan, A. A. 1999. Evaluation of parameters involved in preparation and release of drug loaded in crosslinked matrices of alginate. *Journal of Controlled Release* 57 (3): 223-232.
- ASTM. 1981. Standard test methods for tensile properties of thin plastic sheeting, method D882-80a. Philadelphia, PA.
- Braccini, I. and Pérez, S. 2001. Molecular basis of Ca^{2+} -induced gelation in alginates and pectins: the egg-box model revisited. *biomacromolecules* 2 (4): 1089-1096.
- Chen, C. H. and Lai, L. S. 2008. Mechanical and water vapor barrier properties of tapioca starch/decolorized hsian-tsoa leaf gum films in the presence of plasticizer. *Food Hydrocolloids* 22 (8): 1584-1595.
- Cuq, B., Gontard, N. and Guilbert, S. 1998. Proteins as agricultural polymers for packaging production. *Cereal Chemistry* 75 (1): 1-9.
- Donati, I., Holtan, S., Mørch, Y. A., Borgogna, M. and Dentini, M. 2005. New hypothesis on the role of alternating sequences in calcium-alginate gels. *Biomacromolecules* 6 (2): 1031-1040.
- Dong, Z., Wang, Q. and Du, Y. 2006. Alginate/gelatin blend films and their properties for drug controlled release. *Journal of Membrane Science* 280 (1-2): 37-44.
- Famá, L., Rojas, A. M., Goyanes, S. and Gerschenson, L. 2005. Mechanical properties of tapioca-starch edible films containing sorbates. *LWT - Food Science and Technology* 38 (6): 631-639.
- Fang, Y., Al-Assaf, S., Phillips, G. O., Nishinari, K., Funami, T. and Williams, P. A. 2008. Binding behavior of calcium to polyuronates: Comparison of pectin with alginate. *Carbohydrate Polymers* 72 (2): 334-341.
- Glicksman, J. 1982. Food applications of gums. In Lineback, D. R. and Inglett, G. E. (Eds). *Food Carbohydrate*, p. 270-295. Westport, CT: AVI Publishing Co. Inc.
- Grant, G. T., Morris, E. R., Rees, D. A. and Smith, P. J. C. 1973. Biological interactions between polysaccharides and divalent cations: The egg-box model. *FEBS Letters* 32: 195-198.
- Kester, J. J. and Fennema, O. R. 1986. Edible films and coatings: A review. *Food Technology* 40 (12): 47-59.
- King, A. H. 1982. Brown seaweed extracts (alginates). In Glicksman, M. (Ed). *Food Hydrocolloids*, 2, p. 115-188. Boca Raton, FL: CRC Press.
- Krochta, J. M. 2002. Proteins as raw materials for films and coatings: definitions, current status and opportunities. In Gennadios, A. (Eds). *Protein-based Films and Coating*, p. 1-32. Boca Raton, FL: CRC Press.
- Krochta, J. M. and Mulder-Johnston, C. D. 1997. Edible and biodegradable polymer films: Challenges and opportunities. *Food Technologist* 51 (2): 61-74.
- Larotonda, F. D. S., Matsui, K. N., Sobral, P. J. A. and Laurindo, J. B. 2005. Hygroscopicity and water vapor permeability of Kraft paper impregnated with starch acetate. *Journal of Food Engineering* 71 (4): 394-402.
- Lourdin, D., Coignard, L., Bizot, H. and Colonna, P. 1997. Influence of equilibrium relative humidity and plasticizer concentration on the water content and glass transition of starch materials. *Polymer* 38 (21): 5401-5406.
- Maizura, M., Fazilah, A., Norziah, M. and Karim, A. 2007. Antibacterial activity and mechanical properties of partially hydrolyzed sago starch-alginate edible film containing lemongrass oil. *Journal of Food Science* 72 (6): C324-C330.
- Mali, S. and Grossmann, M. V. E. 2003. Effects of yam starch films on storability and quality of fresh strawberries (*Fragaria ananassa*). *Journal of Agricultural and Food Chemistry* 51 (24): 7005-7011.
- Myllärinen, P., Partanen, R., Seppälä, J. and Forssell, P. 2002. Effect of glycerol on behaviour of amylose and amylopectin films. *Carbohydrate Polymers* 50 (4): 355-361.
- Nisperos-Carriedo, M. O. 1994. Edible coating and films based on polysaccharides. Nisperos-Carriedo, M. O. In Krochta, J. M., Baldwin, E. A. and Nisperos-Carriedo, M. O. (Eds). *Edible coatings and films to improve food quality*, p. 305-335. Lancaster: Technomic Publishing Company, Inc.
- Olivas, G. I. and Barbosa-Cánovas, G. V. 2008. Alginate-calcium films: Water vapor permeability and mechanical properties as affected by plasticizer and relative humidity. *LWT - Food Science and Technology* 41 (2): 359-366.
- Parra, D. F., Tadini, C. C., Ponce, P. and Lugão, A. B. 2004. Mechanical properties and water vapor transmission in some blends of cassava starch edible films. *Carbohydrate Polymers* 58 (4): 475-481.
- Pavlat, A. F., Gossett, C., Camirand, W. and Robertson, G. H. 1999. Ionomeric Films of Alginic Acid. *Journal Food Science* 64 (1): 61-63.
- Phan The, D., Debeaufort, F., Voilley, A. and Luu, D. 2009. Biopolymer interactions affect the functional properties of edible films based on agar, cassava starch and arabinosyran blends. *Journal of Food Engineering*

- 90 (4): 548-558.
- Rhim, J.-W. 2004. Physical and mechanical properties of water resistant sodium alginate films. *Lebensmittel-Wissenschaft und-Technologie* 37 (3): 323-330.
- Rindlav-Westling, A., Stading, M., Hermansson, A.-M. and Gatenholm, P. 1998. Structure, mechanical and barrier properties of amylose and amylopectin films. *Carbohydrate Polymers* 36 (2-3): 217-224.
- Romero-Bastida, C. A., Bello-Pérez, L. A., García, M. A., Martino, M. N., Solorza-Feria, J. and Zaritzky, N. E. 2005. Physicochemical and microstructural characterization of films prepared by thermal and cold gelatinization from non-conventional sources of starches. *Carbohydrate Polymers* 60 (2): 235-244.
- Rubio, M. R. and Ghaly, E. S. 1994. *In vitro* release of acetaminophen from sodium alginate controlled release pellets. *Drug Development and Industrial Pharmacy* 20 (7): 1239-1251.
- Silva, M. A. d., Bierhalz, A. C. K. and Kieckbusch, T. G. 2009. Alginate and pectin composite films crosslinked with Ca²⁺ ions: Effect of the plasticizer concentration. *Carbohydrate Polymers* 77 (4): 736-742.
- Wolff, I. A., Davis, H. A., Cluskey, J. E. and Gundrum, L. J. 1951. Preparation of films from amylose. *Industrial & Engineering Chemistry* 43: 915-919.
- Wu, Y., Geng, F., Chang, P. R., Yu, J. and Ma, X. 2009. Effect of agar on the microstructure and performance of potato starch film. *Carbohydrate Polymers* 76 (2): 299-304.
- Wu, Y., Weller, C., Hamouz, F., Cuppett, S. and Schnepf, M. 2001. Moisture loss and lipid oxidation for precooked ground-beef patties packaged in edible starch-alginate-based composite films. *Journal of Food Science* 66 (3): 486-493.
- Yotsuyanagi, T., Yoshioka, I., Segi, N. and Ikeda, K. 1991. Acid-induced and calcium induced gelation of alginic acid: Bead formation and pH dependent swelling. *Chem Pharm Bull* 39 (4): 1072-1074.
- Zhou, Y. G., Wang, L. J., Li, D., Yan, P. Y., Li, Y., Shi, J., Chen, X. D. and Mao, Z. H. 2009. Effect of sucrose on dynamic mechanical characteristics of maize and potato starch films. *Carbohydrate Polymers* 76 (2): 239-243.