International Food Research Journal 20(5): 2287-2292 (2013)

Journal homepage: http://www.ifrj.upm.edu.my



Nutritional composition and amino acid profile of a sub-tropical red seaweed Gelidium pusillum collected from St. Martin's Island, Bangladesh

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Article history

Received: 9 February 2013 Received in revised form: 9 March 2013 Accepted: 17 March 2013

Keywords

Red seaweed Gelidium pusillum Proximate composition St. Martin's Island

Abstract

Nutritional fact study has prime importance to make the species edible and commercially viable to the food consumers. The proximate chemical composition and amino acid profile of *Gelidium pusillum* were studied to understand the nutritional status. The red seaweed *Gelidium pusillum* was rich in dietary fibre $(24.74 \pm 1.05\%)$, lipid $(2.16 \pm 0.61\%)$ and ash content $(21.15 \pm 0.74\%)$. The mean protein content $(11.31 \pm 1.02\%)$ DW) was within the range of 10-47% for green and red seaweeds and this range was higher than *Gracilaria cornea* (5.47%) DW), *Gracilaria changgi* (6.90%) DW) and *Eucheuma cottonii* (9.76%) DW). *Gelidium pusillum* was found to contained all the essential amino acids, which accounted for 52.08% of the total amino acids. Tyrosine $(26.2 \text{ mg g}^{-1} \text{ protein})$, methionine $(15.8 \text{ mg g}^{-1} \text{ protein})$ and Lysine $(48.3 \text{ mg g}^{-1} \text{ protein})$ were the limiting amino acid of *Gelidium pusillum*. However, the levels of other essential amino acids were above the FAO/WHO requirement pattern (EAA score ranged from 1.14 to 1.62). Aspartic and glutamic acids constituted a substantial amount of the total amino acids (24.68%) of total amino acid). The result from this study suggested that *Gelidium pusillum* could be utilized as a healthy food item for human consumption.

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Introduction

For the last couple of decades, nutritionists and food scientists have given much more concentrations on nutritional evaluation of edible seaweeds (Ratanaarporn and Chirapart, 2006; Kumari et al., 2010). Most of the studies were focused on red seaweeds due to their higher nutritional value compared to other edible seaweeds (brown and green) (Arasaki and Arasaki, 1983; Wong and Cheung, 2000; Marinho-Soriano et al., 2006). Seaweeds have been using as a part of human diet in China, Japan, Thailand and South Korea for many years (Mabeau and Fleurence, 1993; Wong and Cheung, 2000). Depends on species, some seaweeds are generally suitable for making cool, gelatinous dishes or concoctions (Ito and Hori, 1989; Manivannan et al., 2009). In general, seaweeds are considered as low calorie food item, but rich in vitamin, mineral and dietary fibre (Ito and Hori, 1989). Seaweeds are also utilized as animal feed ingredient, row material for fertilizer and as well as various industrial applications (Mabeau and Fleurence, 1993; Fleurence, 1999; Rupérez, 2002; Kumari et al., 2010).

Approximately 1 million tonnes of wet seaweeds

are annually harvested and extracted to produce hydrocolloids (McHugh, 2003). In addition, species belongs to Gelidium are among the most important agarophytes in the world (Santelices, 1974). About 35 seaweed species are harvested in various regions contributing to 40-50% of the world's annual exploitation of agarophytes (Whyte and Englar, 1981). Some seaweed is used in preparing creams, puddings, bears, wines, canned fishes etc. (Bold and Wynne, 1985). Several studies showed that seaweeds are valuable sources of dietary protein, lipid, fibre, vitamin and some essential minerals (Mabeau and Fleurence, 1993; Darcy-Vrillon, 1993; Fleurence, 1999; Novaczek, 2001; Ortiz et al., 2006). Although, it has always been realized that nutritional fact study has prime importance to make the species edible and commercially viable to the consumers (Wong and Cheung, 2000), the nutritional properties of edible red seaweeds are poorly studied in Bangladesh.

Gelidium, Porphyra, Palmaria, Gracilaria, and Eucheuma are the major edible red seaweeds (McLachlan, 1972), where, Gelidium pusillum is commonly known as "Lohit shoibal" in Bangladesh. This red seaweed species abundantly grows in the inshore water of St. Martin's Island, Cox's Bazar.

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Local people of St. Martin's Island usually do not consume this species, but they collect it from the intertidal water by push net or bamboo stick for their livelihood (Zafar, 2005). About 100 families of St. Martin's Island are engaged in collecting seaweeds. Collected seaweeds are sun dried on the sandy beach and export to Myanmar, Singapore and China (Siddique *et al.*, 2013). Previous study on proximate composition of *Gelidium pusillum* is very scantly and nutritional data on *Gelidium pusillum* is not available in the literature. However, proximate chemical composition and amino acid profile of *Gelidium pusillum* are examined in order to provide more comprehensive nutrient information about this species.

Materials and Methods

Study area and sampling method

Gelidium pusillum has been collected randomly by hand-picking from the St. Martin's Island, Cox's Bazar at the time of low-tide during the month of April - June 2008. The St. Martin's Island is situated in the extreme South-Eastern corner (roughly between 20°34′ - 20°39′ N and 92°18′ - 92°21′ E) of Bangladesh which has naturally protected coral reefs. The average turbidity (Secchi disc) of in-shore waters of St. Martin's Island ranges from 1.5 m to 8.0 m. Water temperature of St. Martin's Island ranged between 22 and 29°C (Tomascik, 1997) and salinity of water fluctuated from 21.0-33.5 PSU throughout the year (Zafar, 2005). For this study three sampling stations has been selected at a distance of 200 m in the Western part of St. Martin's Island (see Figure 1). After collection, fresh samples were taken into plastic jar with ice and brought back to the laboratory immediately. In the laboratory, samples were washed by tape water for several times, then gently brushed and rinsed with distilled water. Finally, samples were dried with paper tissue and frozen at -20°C. The dried seaweeds were powdered manually using mortar and pestle and stored in desiccators until the chemical analysis.

Proximate composition analysis

The proximate chemical composition (protein, carbohydrate, crude lipid, fibre, ash and moisture content) of *Gelidium pusillum* was determined according to the standard method (AOAC, 2000). Protein content was analyzed by the Kjeldahl method. A conversion factor of 6.25 has been used to convert total nitrogen content into crude protein. Carbohydrate content was determined as the weight difference using crude protein, lipid, fibre, moisture and ash content data (James, 1996). Crude lipid of

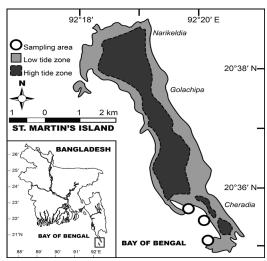


Figure 1. Study area and sampling stations in the St. Martin's Island, Bangladesh

the seaweed was extracted from seaweed powder in a Soxhlet extractor with petroleum ether (Siddique and Aktar, 2011). After ensuring complete extraction of crude lipid, petroleum ether was evaporated and the residue was dried to a constant weight at 105°C. To determine the fibre content of seaweed 2 g samples were boiled with diluted H₂SO₄ (0.3 N). Then the mixture of sample and H₂SO₄ was filtered and washed with 200 ml boiling distilled water and NaOH (0.5 N). After washing by boiling distilled water and acetone the residue was re-extracted and dried at 105°C to constant weight. The moisture content was determined by drying the seaweed samples in an oven at 120°C until a constant weight was obtained and the ash content was estimated by heating the seaweeds in a muffle furnace at 550°C for 4 h (Siddique et al., 2013).

Amino acid analysis

Amino acid analysis was carried out by ionexchange chromatography. A sub-sample (containing 5.0 mg of protein) was taken for acid hydrolysis. 1 ml of HCl acid (6 N) was taken with the sub-sample in a vacuum-sealed hydrolysis vials and heated 22 h at the temperature of 110°C. Norleucine was added to the HCl acid as an internal standard. Few amino acids such as tryptophan, cystine and cysteine were completely lost during the acid hydrolysis process. The tubes were cooled after hydrolysis process and placed in desiccators with some NaOH pellets for 5-6 days. Before analysing the amino acids, the residue was dissolved in a suitable volume of a sample dilution of Na-S buffer with pH 2.2. Then the solution was filtered through a 0.22µm Millipore membrane and prepared for amino acids analysis. A Beckman instrument (model 7300, USA) has been used for the ion-exchange chromatography. During the acid hydrolysis process, some ammonia content

usually comes from the degradation of amino acids (Mosse, 1990; Yeoh and Truong, 1996). Therefore, the ammonia content was included in calculation of protein nitrogen retrieval. The contents of different amino acids recovered are presented as mg g-1 protein and are compared with the FAO/WHO (1991) reference pattern. The essential amino acid (EAA) score was calculated by the following equation:

Essential amino acid score = (mg of EAA in 1 g of test protein / mg of EAA in 1 g of egg protein) x 100

Statistical analysis

All data were expressed in terms of mean ± standard deviation. To estimate the mean percentage and standard deviation, Statistical Package for Social Science (SPSS Version 16.0 for windows) was used in this study.

Results and Discussion

The proximate chemical composition of *Gelidium* pusillum are shown in Table 1. The mean protein content of Gelidium pusillum (11.31 \pm 1.02% DW) was within the range of 10-47% for green and red seaweeds reported by Fleurence (1999). Several studies showed that red seaweed contains higher amount of protein and dietary fibre than that of some other green and brown algae (Arasaki and Arasaki, 1983; Ratana-arporn and Chirapart, 2006). Although, the mean percentage of protein obtained from Gelidium pusillum was lower than some edible red algae; e.g. Gracilaria cervicornis (22.96% DW), Hypnea japonica (19.00%), Hypnea musciformis (18.64% DW) and Porphyra tenera (34.20% DW), but it was higher than *Gracilaria cornea* (5.47% DW), Gracilaria changgi (6.90% DW) and Eucheuma cottonii (9.76% DW) (see Table 2). However, the mean percentage of protein content (11.31 \pm 1.02% DW) recorded from *Gelidium pusillum* is higher than the concentrations found in higher plants (Norziah and Ching, 2000).

Compare to those reported in other edible seaweeds, the mean percentages of crude lipid (2.16) $\pm 0.61\%$ DW) and fibre content (24.74 $\pm 1.05\%$ DW) of Gelidium pusillum was slightly higher (Table 2). Edible seaweeds are not considered as a good source of lipid content as they contain less than 4% of crude lipid at dry weight basis (McDermid and Stuercke, 2003). The mean percentage of crude lipid (2.16 \pm 0.61% DW) obtained from this study is higher than some edible red seaweeds (Gracilaria cervicornis, Gelidium pristoides, Porphyra tenera) reported in previous studies (Arasaki and Arasaki, 1983; Foster and Hodgson, 1998; Marinho-Soriano, 2006) (Table

Table 1. Proximate chemical composition of *Gelidium* pusillum (dry weight basis, n = 9) collected from St.

Martin's Island, Bangladesh				
Nutrient	Gelidium pusillum			
Protein (%)	11.31±1.02			
Crude lipid (%)	2.16 ± 0.61			
Carbohydrate (%)	40.64 ± 2.21			
Fibre (%)	24.74 ± 1.05			
Ash (%)	21.15 ± 0.74			
Moisture (%)	10.85±0.98			
37 - 771	1 1 1			

Note: Values are expressed as mean ± standard

2). Siddique *et al.* (2013) found 1.56% and 1.27% crude lipid in two subtropical red seaweed Hypnea pannosa and Hypnea musciformis, respectively from St. Martin's Island, Bangladesh. However, this result is still lower than the result obtained from Gracilaria changgi (3.30% DW) (Norziah and Ching, 2000). The mean percentage of fibre content ($24.74 \pm 1.05\%$ DW) of Gelidium pusillum is much higher than other red seaweeds (Table 2), but this result is similar with the result found from Gracilaria changgi (24.70%DW) (Norziah and Ching, 2000). The higher amount of crude lipid and fibre in Gelidium pusillum were probably due to the suitable environmental conditions of St. Martin's Island (Haroon et al., 2000). In Gelidium pusillum, the mean percentage of Carbohydrate was $40.64 \pm 2.21\%$ DW. This result concurred well with the previous report on Gelidium pristoides (43.10% DW) and Porphyra tenera (40.70% DW) (Arasaki and Arasaki, 1983; Foster and Hodgson, 1998).

The mean percentage of ash contents (21.15 \pm 0.74%) found in Gelidium pusillum was similar with other red seaweeds. In general, high level of ash was associated with the amount of mineral elements. Previous studies reported that ash content of seaweed varies between 8 and 40% (at dry weight basis) (Mabeau and Fleurence, 1993). The mean percentage of ash found was comparable to those reported in other species i.e., Hypnea japonica (22.10% DW), Hypnea charoides (22.80% DW), Hypnea musciformis (21.57% DW), Gracilaria changgi (22.70% DW) (Norziah and Ching, 2000; Wong and Cheung, 2000; Siddique et al., 2013). Several other studies showed that the variation in ash content depends on seaweed species, geographical origins and their method of mineralization (Nisizawa, 1987; Sanchez-Machado, 2004).

Most of the essential amino acids found in Gelidium pusillum, which accounted for 52.08% of the total amino acid [Level of total EAAs (mg/g of protein)/sum of all measured amino acids (mg/ g protein) x 100%]. The amino acid profiles and the essential amino acid scores of Gelidium pusillum are presented in Table 3. Wong and Cheung (2000) observed that most of the essential amino acids in

Species	Protein	Lipid	Carbohydrate	Fibre	Ash	Moisture	Reference
Gelidium pusillum	11.31	2.16	40.64	24.74	21.15	10.85	Present study
Porphyra tenera	34.20	0.70	40.70	4.80	8.70	-	Arasaki and Arasaki (1983)
Gelidium pristoides	11.80	0.90	43.10	-	14.00	-	Foster and Hodgson (1998)
Eucheuma cottonii	9.76	1.10	26.49	5.91	46.19	10.55	Matanjun et al. (2008)
Gracilaria cervicornis	22.96	0.43	63.12	5.65	7.72	14.33	Marinho-Soriano et al. (2006)
Gracilaria changgi	6.90	3.30	-	24.70	22.70	-	Norziah and Ching (2000)
Gracilaria cornea	5.47	-	36.29	5.21	29.06	-	Robledo and Freile-Pelegrin (1997)
Hypnea pannosa	16.31	1.56	22.89	40.59	18.65	12.35	Siddique et al. (2013)
Hypnea musciformis	18.64	1.27	20.60	37.92	21.57	11.54	Siddique et al. (2013)
Нурпеа јаропіса	19.00	1.42	4.28	53.2	22.10	9.95	Wong and Cheung (2000)
Hypnea charoides	18.40	1.48	7.02	50.30	22.80	10.90	Wong and Cheung (2000)

Table 2. Proximate chemical composition of different red seaweed species reported by various authors (Values are given as percent of dry matter)

Table 3. Amino acid profile (mg g⁻¹ protein) of *Gelidium pusillum*. Values are the average of three determinations and figures in parentheses are the essential amino acids

score

	score	
Amino acids	Gelidium pusillum (Present study)	FAO/WHO (1991) requirement pattern
Argininea	62.7	
Histidinea	4.6	
Isoleucinea	42.1 (1.50)	28
Lysinea	48.3 (0.83)	58
Leucinea	75.2 (1.14)	66
Methionine ^a	15.8 (1.62)	25
Phenyla la nine ^a	31.9	
Tyrosinea	26.2 (0.97)	63
Threoninea	51.5 (1.51)	34
Valinea	44.7 (1.28)	35
Alanineb	51.6	
Aspartic acid ^b	82.2	
Glutamic a cidb	108.8	
Glycine ^b	43.4	
Proline ^b	46.6	
Serine ^b	38.2	
TotalEAA	403	
Totalamino acids (g/100 g DW)	9.8	
	har many and	

Note: aEAA (essential amino acid); bNon-EAA (non essential amino acid)

some subtropical seaweed (*H. japonica*, *H. charoides* and *U. lactuca*), which accounted for 42.1-48.4% of the total amino acids. Furthermore, the levels of all their essential amino acids were comparable to those of the FAO/WHO (1991) requirement pattern. Tyrosine (26.2 mg g⁻¹ protein), methionine (15.8 mg g⁻¹ protein) and Lysine (48.3 mg g⁻¹ protein) were the limiting amino acid of *Gelidium pusillum*. However, the levels of other essential amino acids in this study were above the FAO/WHO (1991) requirement (EAA score ranged from 1.14 to 1.62) (See table 3). With respect to the FAO/WHO (1991) requirement pattern, *Gelidium pusillum* seemed to be able to contribute adequate levels of total EAA for human.

This study revealed that glutamic and aspartic acids are the most abundant amino acids in *Gelidium pusillum*. A number of studies argued that red seaweed contains higher percentages of both aspartic and glutamic acids (Wong and Cheung, 2000; Lourenço, 2002). In *Gelidium pusillum*, aspartic and glutamic acids constituted a substantial amount of the total amino acids (24.68% of total amino acids). Similar results were reported in various other studies

previously (Mabeau *et al.*, 1992; Fleurence, 1999; Wong and Cheung, 2000). In general, most of the seaweeds contain relatively higher amount of free amino acids (Ratana-arporn and Chirapart, 2006). These amino acids provide different types of flavours to several edible seaweeds. Glycine and alanine give a sweet flavour to edible seaweeds (McLachlan, 1972) and aspartic and glutamic acids were responsible for the special flavour and taste of seaweeds (Mabeau *et al.*, 1992).

With the increasing level of education in the developing countries, people are now more concern about nutritional value of consumable food items (Siddique, 2012; Siddique *et al.*, 2012). Since *Gelidium pusillum* have good nutritional value, available and very cheap in south Asian countries, therefore, it could be considered as a low cost healthy food item for human consumption.

Conclusion

With respect to the higher level of crude protein and balanced amino acid profile, *Gelidium pusillum* appeared to be an interesting potential source of plant proteins for human consumption. The higher level of protein, crude lipid and fibre content of this red seaweed species has a great food value from the nutritional and biochemical point of view. The result of this study suggested that *Gelidium pusillum* could be utilized as a healthy food item for human consumption.

Acknowledgement

The authors wish to thank the two anonymous reviewers and the editor in chief Professor Dr. Son Radu for their valuable comments and suggestions for improving this article.

References

AOAC. 2000. Official methods of analysis. 17th edition,

- Association of Official Analytical Chemists, Arlington VA. USA.
- Arasaki, S. And Arasaki, T. 1983. Vegetables from the Sea. Japan Publ. Inc., Tokyo.
- Bold, H.C. and Wynne, M.J. 1985. Introduction to the Algae. Prentice-Hall, Inc., Englewood Cliffs, New Jersey.
- Darcy-Vrillon, B. 1993. Nutritional aspects of the developing use of marine macro-algae for the human food industry. International Journal of Food Science and Nutrition 44: 23-35.
- FAO/WHO. 1991. Protein quality evaluation. Report of joint FAO/WHO expert consultation, Food and Agriculture Organization of United Nations, Rome, Italy.
- Fleurence, J. 1999. Seaweed proteins: biochemical, nutritional aspects and potential uses. Trends in Food Science and Technology 10: 25-28.
- Foster, G.G. and Hodgson, A.N. 1998. Consummation and apparent dry matter digestibility of six intertidal macro-algae by *Turbo sarmaticus* (Mollusca: Vetigastropoda: Turbinidae). Aquaculture 167: 211-227.
- Haroon, A.M., Szaniawska, A., Normant, M. and Janas, U. 2000. The biochemical composition of *Enteromorpha* spp. from the gulf of Gdansk coast on the southern Baltic Sea. Oceanologia 42: 19-28.
- Ito, K. and Hori, K. 1989. Seaweed: Chemical composition and potential foods uses. Food Review International 5: 101-144.
- James, C.S. 1996. Analytical Chemistry of Foods. Chapman and Hall, New York.
- Kumari, P., Kumar, M., Gupta, V., Reddy, C.R.K. and Jha, B. 2010. Tropical marine macroalgae as potential sources of nutritionally important PUFAs. Food Chemistry 120: 749-757.
- Lourenço, S.O., Barbarino, E., De-Paula, J.C., Pereira, L.O.S. and Marquez, U.M.L. 2002. Amino acid composition, protein content and calculation of nitrogen-to-protein conversion factors for 19 tropical seaweeds. Phycological Research 50: 233-241.
- Mabeau, S., Cavaloc, E., Fleurence, J. and Lahaye, M. 1992. New seaweed based ingredients for the food industry. International Food Ingredients 3: 38-45.
- Mabeau, S. and Fleurence, J. 1993. Seaweed in food products: bio-chemical and nutritional aspects. Trends in Food Science and Technology 4: 103-107.
- Manivannan, K., Thirumaran, G., Karthikai, D.G.,
 Anantharaman. P. and Balasubramanian, P. 2009.
 Proximate Composition of Different Group of Seaweeds from Vedalai Coastal Waters (Gulf of Mannar): Southeast Coast of India. Middle-East Journal of Scientific Research 4: 72-77.
- Marinho-Soriano, E., Fonseca, P.C., Carneiro, M.A.A. and Moreira, W.S.C. 2006. Seasonal variation in the chemical composition of two tropical seaweeds. Bioresource Technology 97: 2402-2406.
- Matanjun, P., Mohamed, S., Mustapha, N.M. and Muhammad, K. 2008. Nutrient content of tropical edible seaweeds, *Eucheuma cottonii*, *Caulerpa*

- *lentillifera* and *Sargassum polycystum*. Journal of Applied Phycology 21: 75-80.
- McDermid, K.J. and Stuercke, B. 2003. Nutritional composition of edible Hawaiian seaweeds. Journal of Applied Phycology 15: 513-524.
- McHugh, D.J. 2003. A guide to the seaweed industry. FAO Fisheries Technical Paper, No. 441. Food and Agriculture Organization of United Nations, Rome, Italy.
- McLachlan, J., Craigie, J.S., Chen, L.C.M. and Ogetze, E. 1972. Porphyra linearis Grev: an edible species of Nori from Nova Scotia. Proceedings of the International Seaweed Symposium 7: 473-476.
- Mossé, J. 1990. Nitrogen to protein conversion factor for ten cereals and six legumes or oilseeds: a reappraisal of its definition and determination, variation according to species and to seed protein content. Journal of Agricultural and Food Chemistry 38: 18-24.
- Nisizawa, K., Noda, H., Kikuchi, R. and Watanabe, T. 1987. The main seaweed food in Japan. Hydrobiologia152: 5-29.
- Norziah, M.H. and Ching, C.Y. 2000. Nutritional composition of edible seaweed *Gracilaria changgi*. Food Chemistry 68: 69-76.
- Novaczek, I. 2001. A Guide to the Common Edible and Medicinal Sea Plants of the Pacific Islands, Community Fisheries Training Pacific Series-3A, p. 40. Fiji Island: University of the South Pacific.
- Ortiz, J., Romero, N., Robert, P., Araya, J., Lopez-Hernández, J., Bozzo, C.E., Navarrete, C.E., Osorio, A. and Rios, A. 2006. Dietary fibre, amino acid, fatty acid and tocopherol contents of the edible seaweeds *Ulva lactuca* and *Durvillaea antarctica*. Food Chemistry 99: 98-104.
- Qasim, R. 1991. Amino acids composition of some seaweed. Pakistan Journal of Pharmacological Science 4: 49-54.
- Ratana-arporn, P. and Chirapart, A. 2006. Nutritional evaluation of tropical green seaweeds *Caulerpa lentillifera* and *Ulva reticulata*. Kasetsart Journal of Natural Science 40: 75-83.
- Robledo, D. and Freile-Pelegrin, Y. 1997. Chemical and mineral composition of six potentially edible seaweed species of Yucata'n. Botanica Marina 40: 301-306.
- Rupérez, P. 2002. Mineral content of edible marine seaweeds. Food Chemistry 79: 23-26.
- Sanchez-Machado, D.I., Lopez-Cervantes, J., Lopez-Hernandez, J. and Paseiro-Losada, P. 2004. Fatty acids, total lipid, protein and ash contents of processed edible seaweeds. Food Chemistry 85: 439-444.
- Santelices, B. 1974. Gelidioid algae, a brief resume of the pertinent literature. Marine agronomy. Technical Report, U.S. Sea Grant Program, Hawaii, 111 pp.
- Siddique, M.A.M. 2012. Explaining the role of perceived risk, knowledge, price and cost in dry fish consumption within the Theory of Planned Behaviour. Journal of Global Marketing 25 (4): 181-201.
- Siddique, M.A.M. and Aktar, M. 2011. Changes of Nutritional Value of Three Marine Dry Fishes (*Johnius dussumieri*, *Harpodon nehereus* and *Lepturacanthus*

- savala) during Storage. Food and Nutrition Science 2: 1082-1087.
- Siddique, M.A.M., Aktar, M. and Mohd Khatib, M.A. 2013. Proximate chemical composition and amino acid profile of two red seaweeds (*Hypnea pannosa* and *Hypnea musciformis*) collected from St. Martin's Island, Bangladesh. Journal of Fisheries Sciences.com 7(2): 178-186.
- Siddique, M.A.M., Mojumder, P. andZamal, H. 2012. Proximate composition of three commercially available marine dry fishes (*Harpodon nehereus*, *Johnius dussumieri* and *Lepturacanthus savala*). American Journal of Food Technology 7: 429-436.
- Tomascik, T. 1997. Management Plan for Coral Resources of Narikel Jinjira (St. Martin's Island): Final Report, National Conservation Strategy Implementation Project-1, Ministry of Environment and Forest, Government of Bangladesh.
- Whyte, J.N.C. and Englar, J.R. 1981. The agar component of the red seaweed *Gelidium purpurascens*. Phytochemistry 20: 237-240.
- Wong, K.H. and Cheung, C.K. 2000. Nutritional evaluation of some subtropical red and green seaweeds Part I: proximate composition, amino acid profiles and some physicochemical properties. Food Chemistry 71: 475-482.
- Yeoh, H.H. and Truong, V.D. 1996. Protein contents, amino acid compositions and nitrogen-to-protein conversion factors for cassava roots. Journal of the Science of Food and Agriculture 70: 51-54.
- Zafar, M. 2005. Seaweed Culture in Bangladesh holds promise. Infofish International, Number 1/2005, January/February, 8-10 pp.