

Comparative study of the aqueous and ethanolic extract of *Momordica foetida* on the phenolic content and antioxidant properties

¹*Molehin, O. R. and ²Adefegha, S. A.

¹Department of Biochemistry, Faculty of Science, Ekiti State University Ado-Ekiti P.M.B. 5363 Ado-Ekiti Nigeria

²Functional Food and Nutraceutical Unit, Department of Biochemistry, Federal University of Technology Akure, P.M.B.704 Akure, Nigeria

Article history

Received: 30 August 2013

Received in revised form:

24 September 2013

Accepted: 2 October 2013

Keywords

Momordica foetida

Vegetable

Phenolics

Antioxidant

Medicinal plants

Abstract

The present study was designed to evaluate and compare the phenolic (total phenols and flavonoid) contents and antioxidant activity of aqueous extract of *Momordica foetida* (AEMF) and ethanolic extract of *Momordica foetida* (EEMF) extract of *Momordica foetida* leaves; a popular medicinal plant used for traditional medicine preparations. The antioxidant activity of both extracts was assessed using nitric oxide (NO) radical scavenging and ferric reducing antioxidant power (FRAP) assay. The results revealed that EEMF had significantly ($p < 0.05$) higher total phenol and total flavonoid contents than AEMF. In the same vein, EEMF also exhibited significantly ($p < 0.05$) higher NO radical scavenging ability and ferric reducing antioxidant property than AEMF. The antioxidant properties of the vegetable revealed could be attributed to their phenolic contents. Moreover, EEMF showed better antioxidant activities when compared to AEMF.

© All Rights Reserved

Introduction

Productions of free radicals are essential to any biochemical process and represent an essential part of aerobic life and metabolism. The continuous generation of these radicals is produced by the body's normal use of oxygen such as respiration and some cell mediated immune functions. A dynamic balance exists between the amount of free radicals generated in the body and antioxidants to scavenge them and protect the body against their perilous effects (Roberts *et al.*, 2009; Ramjith *et al.*, 2013). An imbalance between the generation of free radicals and the scavenging abilities of the antioxidants results in oxidative stress. Furthermore, oxidative stress has been implicated in the pathology of many diseases for example arteriosclerosis, cancer, inflammation, neurodegenerative diseases etc (Adesanoye *et al.*, 2012; Goveas and Abraham, 2013).

Several studies have been undertaken on how to delay or prevent the onset of these diseases. The most likely and practical way to fight against degenerative diseases is by boosting body's antioxidant status, which could be achieved by higher consumption of vegetables and fruits which are rich in natural antioxidants, especially phenolics and flavonoids hence the growing interest in research on plants and new plants with antioxidants ability and fit for human consumption (Obboh *et al.*, 2008; Salas *et al.*, 2010).

Therefore, in recent years, considerable attention has been directed towards identification of plants with antioxidant ability that may be used for human consumption (Jain *et al.*, 2009)

Momordica foetida is a medicinal plant that belongs to the family of *Cucurbitaceae*. It is widely distributed in tropical Africa, South-Africa and also found in the Flora of west tropical Africa (Burkill, 1985; Baillie, 2007). The plant has both male and female flowers (Jeffrey, 1990). It has been shown to possess insecticidal properties (Baerts and Lehmann, 2002). The plant is eaten as vegetables in Gabon, Sudan, Uganda, Tanzania and Malawi (Olaniyi, 1975). The leaves have a bitter taste. The plant contains a number of bioactive compounds, including sitosterolglucoside, 5, 25-stigmastadien-3 β -ylglucoside, and 1 β -hydroxyfriedel-6-en-3-one, and several cucurbitane-type triterpenoid derivatives. (Mulholland, 1997). The plants are grazed by cattle and are said to be especially suitable for fattening rabbits. Recent study on the antioxidant potentials of *M. foetida* reveals its scavenging role against lipid peroxidation in rat homogenate (Oloyede and Aluko, 2012). Moreover, there is dearth of information on the antioxidant properties of the polar and non-polar extracts of the plant. Therefore, this study was designed to evaluate and compare the phenolic (total phenols and flavonoid) contents and antioxidant activity of aqueous (AEMF) and ethanolic (EEMF)

*Corresponding author.

Email: molehin.olorunfemi@gmail.com

Tel: +234 803 462 1267

extract of *Momordica foetida* leaves.

Materials and Methods

Sample collection

Fresh samples of *Momordica foetida* leaves were purchased from the market in Otun-Ekiti, Ekiti State Nigeria. The authentication of the leaves was carried out by a Plant Botanist at the Department of Plant Science, Ekiti State University, Ado-Ekiti, Nigeria.

Chemicals and reagents

The following chemicals and reagents: quercetin, gallic acid, Folin–Ciocalteu's reagent, potassium ferricyanide, ferric chloride, potassium acetate, trichloroacetic acid (TCA), methanol, disodium hydrogen phosphate (Na_2HPO_4), sodium dihydrogen phosphate (NaH_2PO_4), sulphanilamide, and sodium carbonate were purchased from Sigma-Aldrich Chemical Co. (St. Louis, MO), Chemie GmbH (Steinheim, Germany) and BDH Chemicals Ltd., (Poole, England).

Aqueous extract preparation

The aqueous extract of the leaves were prepared by soaking the *Momordica foetida* leaves in water for about 24 hours; the mixture were filtered and the filtrate were evaporated under pressure at 40°C. The extracts were stored in the refrigerator for subsequent analysis.

Ethanol extract preparation

The ethanolic extract of the leaves were prepared by soaking the *Momordica foetida* leaves in alcohol for about 24 hours; the mixture were filtered, and the filtrate were evaporated under pressure at 40°C. The extracts were stored in the refrigerator for subsequent analysis

Determination of total phenol content

The total phenol content was determined on the extracts using the method reported by (Singleton *et al.*, 1999). Appropriate dilutions of the extracts were oxidized with 2.5 mL of 10% Folin–Ciocalteu's reagent (v/v) and neutralized by 2.0 mL of 7.5% sodium carbonate. The reaction mixture was incubated for 40 min at 45°C and the absorbance was measured at 765 nm in the spectrophotometer. The total phenol content was subsequently calculated using Gallic acid as standard.

Determination of total flavonoid content

The total flavonoid content of both extracts was determined using a slightly modified method reported by (Meda *et al.*, 2005). Briefly, 0.5 mL of appropriately

diluted sample was mixed with 0.5 mL methanol, 50 μL of 10% AlCl_3 , 50 μL of 1 mol L^{-1} potassium acetate and 1.4 mL water, and allowed to incubate at room temperature for 30 min. Thereafter, the absorbance of the reaction mixture was subsequently measured at 415 nm. The total flavonoid was calculated using quercetin as standard.

Nitric oxide (NO) radical scavenging assay

The scavenging effect of the phenolic extracts on nitric oxide (NO) radical was measured according to the method of (Marcocci *et al.*, 1994). 10 - 400 μl of the phenolic extracts was added in test tubes to 1 ml of sodium nitroprusside solution (25 mM) and tubes incubated at 37°C for 2 hours. An aliquot (0.5 ml) of the incubation solution was removed and diluted with 0.3 ml of Griess reagent (1% sulphanilamide in 5% H_3PO_4 and 0.1% naphthylethylenediaminedihydrochloride). The absorbance of the chromophore formed was immediately read at 570 nm against distilled water as blank with catechin (50 μg) used as standard. Results were expressed as percentage radical scavenging activity (RSA).

Determination of reducing property

The reducing property of the extracts was determined by assessing the ability of the extract to reduce FeCl_3 solution as described by Oyaizu (1986). A 2.5 mL aliquot was mixed with 2.5 mL of 200 mmol L^{-1} sodium phosphate buffer (pH 6.6) and 2.5 mL of 1% potassium ferric cyanide. The mixture was incubated at 50°C for 20 min and then 2.5 mL of 10% trichloroacetic acid was added. This mixture was centrifuged at 650 rpm for 10 min. 5 mL of the supernatant was mixed with an equal volume of water and 1 mL of 0.1% ferric chloride. The absorbance was measured at 700 nm. The ferric reducing antioxidant property was subsequently calculated using ascorbic acid as standard.

Statistical analysis

The result of the triplicate experiments were pooled and expressed as mean \pm standard deviation (SD). Student t-test was carried out to ascertain the level of significance. Significance was accepted at $p \leq 0.05$ (Zar, 1984).

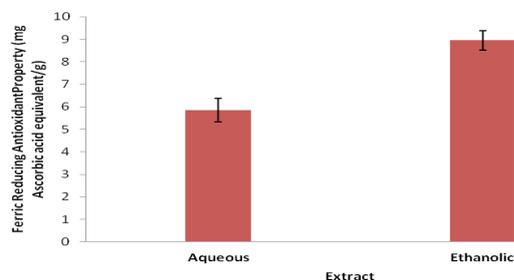
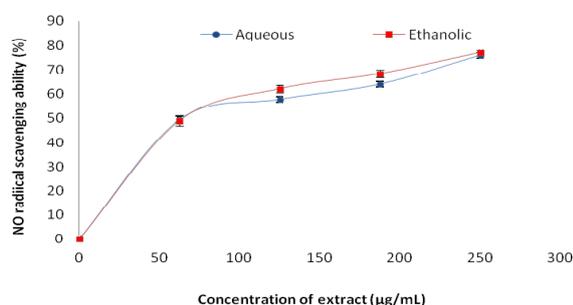
Results

The results of the phenolic content of aqueous extract of *Momordica foetida* (AEMF) and ethanolic extract of *Momordica foetida* (EEMF) are shown in Table 1 and reported as gallic acid equivalent (GAE) for total phenol content and quercetin equivalent

Table 1. Phenolic contents of aqueous (AEMF) and ethanolic (EEMF) extract of *Momordica foetida*

| | Total phenol (mgGAE/g) | Total flavonoids (mgQE/g) |
|----------------|---------------------------|------------------------------|
| AEMF aqueous | 3.93 ± 0.17 ^a | 2.88 ± 0.04 ^c |
| EEMF ethanolic | 5.55 ± 0.46 ^b | 3.94 ± 0.07 ^d |

Values represent mean ± standard deviation of triplicate readings, n = 3. Values with different superscript letters along the same column are significantly (p < 0.05) different

Figure 1. Ferric reducing antioxidant property of ethanolic and aqueous extract of *Momordica foetida*Figure 2. Nitric oxide radical scavenging ability of ethanolic and aqueous extract of *Momordica foetida*

antioxidant capacity (QEAC) for total flavonoid content. The result revealed that there is significant ($P < 0.05$) difference in total phenol contents of AEMF (3.93 ± 0.17 mg/g) and EEMF (5.55 ± 0.46 mg/g). Likewise, the total flavonoid contents of EEMF extract (3.94 ± 0.07 mg/g) was significantly higher ($P < 0.05$) than AEMF (2.88 ± 0.04 mg/g). The *in vitro* antioxidant indices (ferric reducing antioxidant property and nitric oxide scavenging property) of both extracts were presented in Figure 1 and 2 respectively. The trend of the result of the total phenol content of both extracts agrees with that of the total flavonoid content in that EEMF showed higher phenolic content than AEMF. Furthermore, the ferric reducing antioxidant property of the extracts was assessed for their ability to reduce Fe (III) to Fe (II) and expressed as ascorbic acid equivalent (AAE), as shown in Figure 1. The result revealed that both extracts possess good reducing potentials. However, Moreover, EEMF (8.95 mgAAE/g) had significant difference ($p < 0.05$) higher reducing power than AEMF (5.85 mgAAE/g). Also, the NO radical scavenging ability of both extracts is presented in Figure 2. Judging by the IC_{50} (extract concentration that caused 50% inhibition), the result revealed that there was no significant difference between EEMF

($IC_{50} = 64.67$ µg/ml) and AEMF ($IC_{50} = 65.33$ µg/ml). It is noteworthy that the phenolic contents of both extracts are consistent with their antioxidant properties in that EEMF exhibited higher phenolic contents and antioxidant properties than AEMF.

Discussion

Strong research evidences has shown that most of the biological actions of plant foods are linked to their phenolic contents (Scalbert *et al.*, 2005; Oboh and Rocha, 2007). Phenolic compounds exert their antioxidant properties by scavenging free radicals, chelating metals, reducing action and inhibition of lipid peroxidation (Alia *et al.*, 2003).

Total phenolic compounds are reported as milligram gallic acid equivalents/g equivalents (mgGAE/g) as shown in Table 1. The total phenolic contents of AEMF and EEMF were 3.93 ± 0.01 and 5.55 ± 0.46 mgGAE/g of sample, respectively. The total flavonoid contents of AEMF and EEMF were 2.88 ± 0.04 and 3.94 ± 0.07 mg quercetin equivalent/g of sample. EEMF had higher total phenol and flavonoid contents than AEMF ($P < 0.05$). It has been reported that green leafy vegetables, soft fruits and medicinal plants exhibited higher levels of flavonoid (Sultan and Anwar, 2009). Phenols and polyphenolic compounds, such as flavonoids, are widely found in food products derived from plant sources, and they have been shown to possess significant antioxidant activities (Van Acker *et al.*, 1986).

The ability of the extracts to reduce ferric ion to ferrous ion was determined and the result was presented as ascorbic acid equivalent. The result revealed that EEMF was found to be significantly higher ($p < 0.05$) as compared to AEMF (Figure 1) Since FRAP assay is easily reproducible and linearly related to molar concentration of the antioxidant present, it can be reported that AEMF and EEMF may act as free radical scavenger, capable of transforming reactive free radical species into stable non radical products.

Figure 2 shows the Nitric oxide radical scavenging ability of ethanolic and aqueous extract of *Momordica Foetida*. The extract effectively reduced the generation of nitric oxide from sodium nitroprusside (Figure 2). Inhibition of nitric oxide radical is a measure of antioxidant activity of plant extracts. Scavenging of nitric oxide radical is based on the generation of nitric oxide from sodium nitroprusside in buffered saline, which reacts with oxygen to produce nitrite ions that can be measured by using Griess reagent (Marcocci *et al.*, 1994). Both AEMF and EEMF decreased the amount of

nitrite generated from the decomposition of sodium nitroprusside which indicated the presence of antioxidant principles in the extract. The percentage scavenging activity increased with increasing concentration of the extract. Moreover, there was no significant difference ($p \geq 0.05$) in the Nitric oxide radical scavenging ability of AEMF and EEMF.

Conclusion

In this study, we have demonstrated that both ethanolic and aqueous extract of *Momordica foetida* exhibited antioxidants properties. The antioxidant properties of the vegetable revealed could attributed to their phenolic contents. Moreover, Ethanolic extract of *Momordica foetida* showed better antioxidant activities when compared to aqueous extract of *Momordica foetida*.

References

- Adesanoye, O.A., Molehin, O.R., Delima, A.A., Adefegha S.A. and Farombi, E.O. 2012. Modulatory effect of methanolic extract of *Vernonia amygdalina* (MEVA) on tert-butyl hydroperoxide-induced erythrocyte haemolysis Cell Biochemistry and Function (wileyonlinelibrary.com) DOI: 10.1002/cbf.2933
- Alia, M., Horcajo, C., Bravo, L. and Goya, L. 2003. Effect of grape antioxidant dietary fiber on the total antioxidant capacity and the activity of liver antioxidant enzymes in rats. Nutritional Research 23: 1251–1267
- Baerts, M. and Lehmann, J. 2002. *Momordica foetida* *prelude* medicinal plants database 8: 74-80
- Baillie, J.K., Bates, M. G. D., Thompson, A. A. R., Waring, W. S., Partridge, R. W., Schnopp, M. F., Simpson, A., and Gulliver-Sloan, F. 2007. Endogenous urate production Augments Plasma Antioxidant Capacity in Healthy Lowland Subjects exposed to High Altitude. Chest 131(5):1473–1478.
- Burkill, H.M. 1985. The useful plants of west tropical Africa 2nd edition 1, 960
- Goveas, S.W. and Abraham, A. 2013 Evaluation of antimicrobial and antioxidant activity of stem and leaf extracts of *Coscinium fenestratum*. Asian Journal of Pharmaceutical and Clinical Research 6 (3): 218-221
- Gupta, A. D., Pundeer, V., Bande, G., Dhar, S., Ranganath, I. R. and Kumari G. S. 2009. Evaluation of antioxidant activity of four folk antidiabetic medicinal plants of India Pharmacology online 1: 200-208
- Jain, S., Gupta, A., Malviya, N. and Suhur, H.2009. Comparative antioxidant potential screening of polyherbal formulations. Advances in Pharmacology and Toxicology 10 (1): 101-110.
- Jeffrey, C. 1990. *Cucurbitaceae* flora of tropical East Africa pp 157
- Marcocci, P.L., Sc kaki, A. and Albert, G.M.1994. Antioxidant action of *Ginkgo biloba* extracts EGP 761. Methods in Enzymology 234: 462-475
- Meda, A, Lamien, C.E., Romito, M., Millogo, J. and Nacoulma, O.G. 2005. Determination of the total phenolic, flavonoid and praline contents in Burkina Fasan honey, as well as their radical scavenging activity. Food Chemistry 91:571 –577
- Mulholland, D. A., Sewram, V., Osborne, R., Pegel, K. H. and Connolly, J. D. 1997. *Cucurbitane triterpenoids* from the leaves of *Momordica foetida* Phytochemistry 45 (2): 391 – 395
- Olaniyi, A. A. 1975. A neutral constituent of *Momordica foetida*. Llydia 38(4): 361-362
- Oboh, G, and Rocha, J.B.T 2007. Antioxidant in Foods: A New Challenge for Food processors. Leading Edge Antioxidants Research, Nova Science Publishers Inc. New York US, 35- 64
- Oboh, G; Raddatz, H and Henle. T. 2008. Antioxidant properties of polar and non-polar extracts of some tropical green leafy vegetables Journal of Science and Food Agriculture 88: 2486–2492
- Oloyede, O.I. and Aluko, O.M. 2012. Determination of Antioxidant Potential of *Momordica foetida* Leaf Extract on Tissue Homogenate Science Journal of Medicine and Clinical Trials. doi: 10.7237/sjmct/225
- Oyaizu, M. 1986. Studies on products of browning reaction: antioxidative activity of products of browning reaction prepared from glucosamine. Japan Journal of Nutrition 44: 307–315
- Ramjith, U.S., Roopitha, B. and Jacob, C.M. 2013. Isolation anti-diabetic and antioxidant evaluation of aqueous extract of *Cansjera rheedii* leaves. Asian Journal of Pharmaceutical and Clinical Research 6(3): 228-234
- Roberts, R. A., Laskin, D. L., Smith, C. V., Robertson, F. M., Allen, E. M. G., Doorn, J. A. and Slikkerk, W. 2009. Nitrate and oxidative stress in toxicology and disease. Toxicological Sciences 112: 4–16
- Salas, P.G., Soto, A.M., Carretero, A. S. and Gutiérrez, A. F. 2010. Phenolic Compound-Extraction Systems for Fruit and Vegetable Samples. Molecules 15: 8813-8826
- Scalbert, A., Manach, C., Morand, C. and Remesy, C. 2005. Dietary polyphenols and the prevention of diseases. Critical Reviews in Food Science and Nutrition 45:287–306
- Singleton, V.L., Orthofer, R. and Lamuela-Raventos, R.M.1999. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin–Ciocalteu's reagent. Methods in Enzymology 299:152–178
- Sultan, B. and Anwar, F. 2009. Flavonols (Kampeferol, quercetin, myricetin) contents of selected fruits, vegetables and medicinal plants. Food Chemistry 108(3):879-884
- Van Acker, S.A.B.E., Van Den Berg, D.J., Tromp, M.N.J.L., Griffioen, D.H., Van Bennekom, W.P. and Van der

Vijgh, W.J.F. 1996. Structural aspects of antioxidant activity of flavanoids. *Free Radical Biology and Medicine* 20(30):331-342

Zar, J.H. 1984. *Biostatistical Analysis*. USA: Prentice-Hall, Inc 620.