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# Optimization of essential oil and fucoxanthin extraction from *Sargassum binderi* by Supercritical Carbon Dioxide (SC-CO<sub>2</sub>) extraction with ethanol as co-solvent Using Response Surface Methodology (RSM)

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# <u>Keywords</u>

Fucoxanthin Supercritical carbon dioxide Response surface methodology Sargassum binderi conventional solvent extraction as it is less toxic, less hazardous to the environment and preserves the bioactivity of fucoxanthin. A face-centered central composite design (FCCCD) based on response surface methodology (RSM) was employed for SC-CO, extraction of oils and fucoxanthin from the brown seaweed Sargassum binderi, with ethanol as a co-solvent. Three independent parameters namely, extraction temperature (A: 40, 50, 60°C), pressure (B: 2900, 3625, 4350 psig and particle size (C: 90, 500 and 1000 μm) were investigated to optimize extraction oil yields (EOY) and fucoxanthin yields (FY). A regression model was developed, tested for quality of fit  $(R^2)$  and expressed in the form of 3D response surface curve and 2D contour. The optimum extraction conditions were obtained at extraction temperature (A) 50°C, pressure (B) 3625 psig and particle size (C) 500  $\mu$ m. Under these conditions, optimal EOY and FY were 10.04 mg/g and 3188.99  $\mu$ g/g, respectively. The difference between the lowest and the highest response in EOY and FY were 5.44 - 10.04 mg/g and  $2109.10 - 3188.90 \mu$ g/g, respectively. The lowest yields were identified at 60°C, 2900 psig and 1000 µm. The regression models generated showing interactions between the variables and EOY and FY response were significant as tested by ANOVA (p < 0.0005 and p < 0.0007, respectively) with high R<sup>2</sup> values (0.9848 and 0.9829, respectively). Interactions between the parameters had a strong synergistic effect on EOY and FY values, as indicated by the 3D response surface curve and 2D contour. The experimental results matched the predicted results closely. This indicated the suitability of the models developed and the success of FCCCD under RSM in optimizing the S. binderi extraction conditions.

Supercritical carbon dioxide (SC-CO<sub>2</sub>) extraction of fucoxanthin is more advantageous over

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# Introduction

In an earlier study, reversed phase-HPLC (RP-HPLC) equipment was successfully used to quantify fucoxanthin content from three Malaysian brown seaweeds, *Sargassum binderi, S. duplicatum* and *Padina australis* (Noviendri *et al.*, 2011; Jaswir *et* 

<u>Abstract</u>

*al.*, 2011). The fatty acid contents were analyzed by gas chromatography in the form of fatty acid methyl esters (FAMEs). *S. binderi* is a good candidate for fatty acid sources from seaweeds because the n6/n3 ratio is less than 1 (0.87) (Noviendri *et al.*, 2011), Contributed by high n-3 and low n-6. The n6/n3 ratio is beneficial for health due to several sources

of information suggesting that human beings evolved a diet in which the ratio of n-6 to n-3 essential fatty acids (EFAs) was about 1 (Simopoulus, 2002).

Fucoxanthin is a major marine carotenoid, found in edible seaweeds (Roh *et al.*, 2008). This compound is one of the most abundant carotenoids in nature (Matsuno, 2011; Miyashita, 2008), especially in the marine environment (Dembitsky and Maoka, 2007).

Currently, the most common method for fucoxanthin extraction is by liquid solvent extraction such as acetone, ethanol and dimethyl sulfoxide (DMSO). Fucoxanthin has been extracted from the microalgae *Phaeodactylum tricornutum* using ethanol (Kim et al., 2012), the brown seaweed Laminaria japonica using DMSO (Wang et al., 2005) and ethanol (Zhang et al., 2008), Undaria pinnatifida using acetone (Sugawara et al., 2006) and chloroform/ methanol (Maeda et al., 2008), and Hijikia fusiformis (Yan et al., 1999), Eisenia bicyclis, respectively using acetone (Kim et al., 2011). However, conventional techniques can damage the functional properties of the extracts and are also potentially hazardous to the environment (Foster et al., 1993). Supercritical fluid extraction (SFE) eliminates the disadvantages of conventional technique which leads to degradation of thermally labile compounds (Roh et al., 2006), and leaves traces of toxic solvents in the solute (Döker et al., 2010).

SFE technology has been used in large-scale extraction of some essential oils (Abbas et al., 2008) and fucoxanthin from Undaria pinnatifida (Roh et al., 2008). The most commonly used SFE is supercritical carbon dioxide (SC-CO<sub>2</sub>) extraction (Santoyo et al., 2006; Machmudah et al., 2007), because it has a favorable critical pressure (Pc =1071 psig) and critical temperature (Tc =  $31.1^{\circ}$ C) (Lang et al., 2001; Machmudah et al., 2007; Romo- Hualde et al., 2012). SC-CO<sub>2</sub> extraction exhibits good density, high diffusivity, low surface tension and low viscosity, which play key roles in enabling the solvent to readily penetrate the solid biomass matrix as well as in extracting the solutes (Muthukumaran et al., 1999; Baek et al., 2004; Roh et al., 2008). SC-CO, extraction is also chemically inert under many conditions, non-flammable, nontoxic, and inexpensive (Lang et al., 2001; Turner and Mathiasson, 2001; Machmudah et al., 2007; Roh et al., 2008; Abbas et al., 2008; Norulaini et al., 2009; Romo- Hualde et al., 2012). These properties of SC-CO<sub>2</sub> make the products more advantageous in the field of food, pharmaceutical (Abbas et al., 2008) or medicinal extract (Doker et al., 2010), and cosmetics (Machmudah et al., 2007).

Therefore, the purpose of this study was to

optimize essential oil and fucoxanthin extraction from selected Malaysian brown seaweed *(S. binderi)* by SC-CO<sub>2</sub> extraction with ethanol as co-solvent under various: temperatures, pressures and particle sizes of the sample using response surface methodology (RSM).

### Materials and methods

# Materials

Plant material (brown seaweed) used in this study was *S. binderi*. It was freshly collected from Port Dickson, Negeri Sembilan, Malaysia in August 2011. Food grade carbon dioxide (99.9% pure) was used and ethanol as co-solvent as well as other reagents were of analytical grade.

#### Sample preparation

Fresh brown seaweed *(S. binderi)* was washed thoroughly with fresh water to remove salt and sand attached to the surface (Heo and Jeon, 2009). The cleaned sample was frozen at -80°C and then dried in freeze-drier for 3 days (Roh *et al.*, 2008). The dried *S. binderi* was ground in a mill before treatment through a mesh sieve with sizes 90, 500 and 1000 μm.

#### Experimental design

RSM was used to optimize the conditions that enhanced oil and fucoxanthin extraction by SC-CO<sub>2</sub> extraction with ethanol as co-solvent. A face-centered central composite design (FCCCD) under RSM developed by the Design Expert software (Version 6.0.8, Stat-Ease Inc., Minneapolis, USA) (Muntari *et al.*, 2012) was used to optimize the three significant extraction conditions: pressure, temperature and sample particle size, to yield of oil and fucoxanthin extract.

A set of 15 experimental runs with one center point (Run 13) was generated. Subsequently, three different levels, low (-1), medium (0) and high (+1) were used to study the independent variables. The extracted oil and fucoxanthin yield were considered as the response ( $Y_1$ ) and ( $Y_2$ ), respectively. The following second-order polynomial equation explains the relationship between dependent and independent variables (Muntari *et al.*, 2012):

 $Y_{1} \text{ or } Y_{2} = \beta 0 + \beta 1A + \beta 2B + \beta 3C + \beta_{11}A^{2} + \beta_{22}B^{2} + \beta_{33}C^{2} + \beta_{12}AB + \beta_{13}AC + \beta_{23}BC....(1)$ 

Where  $Y_1$  is the dependent variable (extracted oil yield, EOY), and  $Y_2$  is the dependent variable (fucoxanthin yield, FY); A, B and C are independent variables (temperature, pressure and particle size of

sample, respectively);  $\beta_0$  is an intercept term;  $\beta_1$ ,  $\beta_2$ and  $\beta_3$  are linear coefficients;  $\beta_{12}$ ,  $\beta_{13}$  and  $\beta_{23}$  are the interaction coefficients; and  $\beta_{11}$ ,  $\beta_{22}$ , and  $\beta_{33}$  are the quadratic coefficients.

The developed regression model was evaluated by analyzing the values of regression coefficients, analysis of variance (ANOVA), p- and F-values (Muntari *et al.*, 2012). Then, the quality of fit of the polynomial model equation was expressed by the coefficient of determination,  $R^2$  (Bari *et al.*, 2009; Muntari *et al.*, 2012). Furthermore, to explain the relationship between the experimental levels of each of variables under study and the responses, the fitted polynomial equation was expressed in the form of 3D response surface and contour (Muntari *et al.*, 2012).

Determination of extracted oil and fucoxanthin yield The estimated yield of oil obtained from S. binderi by SC-CO, with ethanol as co-solvent was calculated directly as a ratio between the weight of oil obtained from extraction process and the weight of dried sample used in this optimization, in percentage. The determination of carotenoid (fucoxanthin) yield was adopted from the method of Gu et al. (2008) and Chen et al. (2006) with slight modifications. Briefly, the extract of S binderi was diluted with acetone and the solution was transferred to CELLSTAR<sup>®</sup> 96 well flat bottom plates (Greiner Bio-one) (Mori et al., 2004) and measured by microplate reader (Tekan/ infinite M200, NanoQuan) at 450 nm (maximum wavelength for detecting fucoxanthin) (Yan et al., 1999; Simopoulus, 2002; Mori et al., 2004; Maeda et al., 2006; Nakazawa et al., 2009; Noviendri et al., 2011a, 2011b; Jaswir et al., 2011; Jaswir et al., 2013). The fucoxanthin yield (FY) ( $\mu g/g$  dried weight sample) was calculated according to the following formula (Chen et al., 2006):

FY ( $\mu$ g/g dried weight sample) = 1000ADV/0.16W .....(2)

Where A is the absorbance value of diluted extraction at 450 nm, D is the dilution ratio, V is the volume of the acetone. 0.16 is extinction coefficient of carotenoids (fucoxanthin), and W (g) is the weight of dried *S. binderi*.

# **Results and discussion**

## Optimization of SC-CO, extraction condition by RSM

The use of statistical experimental design is a vital tool in optimizing conditions (Muntari *et al.*, 2012) that cause an increment of several folds in oil

Table 1. An FCCCD of three independent variables with their actual and coded values and one center point showing the experimental and predicted response.

Run	Т	Р	PS	EOY (mg/g dw)		FY (µg/g <i>dw</i> )	
	(°C)	(psig)	(µm)	Ex.	Pr.*	Ex.	Pr.**
1	60	2900	1000	5.44	5.46	2109.10	2109.30
	(+1)	(-1)	(+1)				
2	50	3625	1000	7.78	7.77	2600.30	2637.68
	(0)	(0)	(+1)				
3	40	4350	90	7.23	7.19	2589.40	2577.28
	(-1)	(+1)	(-1)				
4	60	3625	500	8.87	9.12	3000.10	3025.69
	(+1)	(0)	(+1)				
5	50	2900	500	8.51	8.72	3011.10	2942.32
	(0)	(-1)	(0)				
6	60	4350	90	7.91	7.93	2885.70	2860.28
	(+1)	(+1)	(-1)				
7	40	2900	1000	5.67	5.60	2300.00	2312.81
	(-1)	(-1)	(+1)				
8	40	2900	90	7.22	7.26	2579.10	2596.19
	(-1)	(-1)	(-1)				
9	60	4350	1000	7.89	7.81	2645.90	2623.85
	(+1)	(+)	(+1)				
10	50	4350	500	9.79	9.76	3029.30	3167.23
	(0)	(+1)	(0)				
11	50	3625	90	8.47	8.66	2988.90	2897.58
	(0)	(0)	(-1)				
12	40	4350	1000	7.39	7.53	2785.50	2757.16
	(-1)	(+1)	(+1)				
13	50	3625	500	10.04	9.69	3188.90	3118.60
	(0)	(0)	(0)				
14	40	3625	500	8.87	8.80	2954.80	2965.36
	(-1)	(0)	(0)				
15	60	2900	500	7.78	7.58	2786.30	2808.98
	(+1)	(-1)	(0)				

T: temperature; P: pressure; PS: particle size; EOY: extracted oil yield; FY: fucoxanthin yield; Ex.: experiment; Pr.: predicted. \*Second order polynomial (Eqn 3) was used to estimate the predicted response (extracted oil). \*\* Second order polynomial (Eqn. 4) was used to estimate the predicted response (carotenoid yield).

and carotenoid (fucoxanthin) yields. Wu *et al.* (2007) reported that the main advantage of RSM is to reduce the number of experimental trials needed to evaluate multiple variables and interactions. In addition, RSM is less laborious and time-consuming compared to other approaches (Hossain *et al.*, 2011; Salihu *et al.*, 2011).

In this study, an FCCCD under RSM was used to determine the optimal conditions of the three significant factors (temperature, pressure and particle

Table 2. ANOVA	of a quad	lratic model	for EOY.
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Source	Sum of square	F-value	p-value
Model	22.61	36.01	0.0005
Temperature,	0.22	3.17	0.1349
A			
Pressure, B	3.24	46.46	0.0010
Particle size,	1.98	29.39	0.0031
С			
A <sup>2</sup>	1.34	19.24	00071
B"	0.52	7.24	0.0416
C <sup>2</sup>	5.21	74.67	0.0003
AB	0.087	1.25	0.3140
AC	0.11	1.52	0.2723
BC	2.02	29.98	0.0030

 $\overline{R}^2 = 0.9848$ , Adjusted  $\overline{R}^2 = 0.9575$ , Adequate precision = 19.934, p < 0.05 was considered to be significant.

size of the sample) to yield oil and fucoxanthin extract. For each run, the experimental (observed) results along with the predicted EOY and FY obtained from the regression equations for the 15 combinations are shown in Table 1.

In this study, a co-solvent ethanol was used in SC- $CO_2$  to extract the semi-polar carotenoid fucoxanthin. In the SC- $CO_2$  extraction of carotenoid from a natural source, the use of co-solvents such as ethanol (Nobre *et al.*, 2006) has been used to improve the extraction efficiency (Naranjo- Mdad *et al.*, 2000; Vassapollo *et al.*, 2004). The presence of a polar co-solvent can increase the solubility of polar compounds and the selectivity of the process (Abbas *et al.*, 2008) is due to the polar character of carotenoids. The formation of hydrogen bonds with ethanol present in the  $CO_2$  stream and swelling of the biomass pore facilitates the release of the pigments from the samples (Nobre *et al.*, 2006).

From this study, the results demonstrated that optimal extracted oil yield (EOY) and fucoxanthin yield (FY) by SC-CO<sub>2</sub> extraction with ethanol as cosolvent are 10.04 mg/g and 3188.99  $\mu$ g/g, respectively (run 13, Table 1) achieved at 50°C, 3625 psig and 500  $\mu$ m. Furthermore, the lowest amounts were observed in run 1 (5.44 mg/g and 2109.10  $\mu$ g/g), where the factors such as temperature and particle size of the sample were at highest conditions (60°C and 1000  $\mu$ m, respectively), whereas the pressure was at lowest condition (2900 psig). This study shows that the design matrix of FCCCD further improved the EOY and FY, such that the difference between the lowest and the highest response (5.44 – 10.04 mg/g; 2109.10 - 3188.90  $\mu$ g/g), respectively.

Furthermore, a second order regression equation

Table 3. ANOVA of a quadratic model for l
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Source	Sum of square	F-value	<i>p</i> -value
Model	1.189E+006	32.01	0.0007
Temperature,	3828.07	0.93	0.3798
Α			
Pressure, B	1.582E+005	38.35	0.0016
Particle size,	1.740E+005	42.18	0.0013
С			
A <sup>z</sup>	38948.84	9.44	0.0277
B"	10474.14	2.54	0.1720
C <sup>2</sup>	2.932E+005	71.07	0.0004
AB	2462.02	0.60	04746
AC	86824.97	21.04	0.0059
BC	1.075E+005	26.06	0.0038

 $R^2 = 0.9829$ , Adjusted  $R^2 = 0.9522$ , Adequate precision = 20.171, p < 0.05 was considered to be significant.

showed the dependence of EOY and FY on SC- $CO_2$  extraction components. The parameters of the equation were obtained by multiple regression analysis of the experimental data (Salihu *et al.*, 2011). An empirical relationship between the screened variables and response were expressed in terms of the second-order polynomial equation:

 $Y_{1}(EOY, mg/g dried weight) = +9.66 + 0.15A + 0.57B - 045C - 0.72A^{2} - 0.45B^{2} - 1.44C^{2} + 0.10AB - 0.12AC + 0.50BC \dots (3)$ 

Where the EOY is the response  $(Y_1)$  and A, B and C are temperature, pressure, and particle size of the sample, respectively.

Where the FY is the response  $(Y_2)$  and A, B and C are the temperature, pressure, and particle size of the sample, respectively.

The adequacy of the model for EOY and FY were checked using ANOVA which was tested using statistical analysis of Fisher and the results are shown in Table 2 and Table 3. For EOY (Table 2), the model F value of 36.01 and p-value of < 0.0005 imply that the model is significant, suggesting that there is only 0.05% chance that the model F value could occur due to noise. Model terms with Probability > F (less than 0.05) are considered significant, while those greater than 0.10 are insignificant (Saliu *et al.*, 2011).

Furthermore, for FY (Table 3), the F value of



Figure 1. 3D response surface curves and 2D contour of the combined effects of temperature, pressure, the particle size of the sample on EOY by SC-CO<sub>2</sub> extraction with ethanol as cosolvent. Temperature and pressure at fixed level of particle size of sample (A), temperature and particle size at fixed level of pressure (B), pressure and particle size of the sample at fixed level of temperature (C).

32.01 and p-value of < 0.0007 imply that the model is significant, suggesting that there is only 0.07% chance that the model F value could occur due to noise. Bari *et al.* (2009) have reported that a greater F-value indicates that the factors adequately explain the variation in data about its mean, and estimated factor effects are real. From this study, the model terms with Probability > F (less than 0.05) are considered significant.

The  $R^2$  value closer to 1 denotes a better correlation between the experiment (observed) and predicted values (Salihu *et al.*, 2011). For EOY (Table 2), the higher values of  $R^2$  (0.9848) and adjusted  $R^2$ (0.9829) also indicated the efficacy of the model and 98.48% or 98.29% of variations could be accounted for by model equation. Thus, for a good statistical model, the  $R^2$  value should be in the range of 0 – 1.0, and the closer the value is to 1.0, the more fit the model is deemed to be.

Moreover, adequate precision measures signal



Figure 2. 3D response surface curves and 2D contour of the combined effects of temperature, pressure, the particle size of the sample on FY by SC-CO<sub>2</sub> extraction with ethanol as cosolvent. Temperature and pressure at fixed level of particle size of sample (A), temperature and particle size at fixed level of pressure (B), pressure and particle size of the sample at fixed level of temperature (C).

to noise ratio and a value of >4 is considered a prerequisite for desirable models. The adequate precision value of 19.937 for EOY indicates that the model can be used to navigate the design space. Furthermore, for FY (Table 3), the higher values of  $R^2$  (0.9829) and adjusted  $R^2$  (0.9522) also indicated the efficacy of the model and 98.29% or 95.22% of variations could be accounted for by model equation. The adequate precision value of 20.171 for FY also indicates that the model can be used to navigate the design space due to the adequate precision measures signal to noise ratio and a value >4. Thus, this is considered a prerequisite for desirable models.

The coefficient values of regression equation are listed below in Table 2 and 3. The p-value is used as a tool to check the significance of each coefficient, which also indicates the interaction strength between each independent variable (Li *et al.*, 2012). The smaller the p-value, the larger the significance of the corresponding coefficient (Li *et al.*, 2012).

For EOY (Table 2), the responses revealed that (B – pressure) and (C – particle size of sample), one interaction terms BC (pressure and particle size of sample), and all the quadratic coefficients (A<sup>2</sup>, B<sup>2</sup> and C<sup>2</sup>) were significant (p < 0.05) and had remarkable effects on the overall extracted oil. Similarly, for FY (Table 3), the responses revealed that (B – pressure) and (C – particle size of the sample) were significant (p < 0.05). However, for FY, there are two interaction terms AC (temperature and particle size of the sample) and BC (pressure and particle size of the sample), and the two quadratic coefficients (A<sup>2</sup> and C<sup>2</sup>) were significant (p < 0.05) that had remarkable effects on the overall FY.

Tanyldizi et al. (2005) reported that the 3D response surface and 2D contour plots are the graphical representation of the regression equation used to determine the optimum values of the variables within the ranges considered. The 3D response surface and 2D contour plots of the combined effects of temperature, pressure and particle size of the sample for EOY and FY by SC-CO<sub>2</sub> extraction with ethanol as co-solvent are shown in Figures 1 and 2, respectively. The 3D plots are based on the function of the condition of two variables with the other variable being at its optimum level. The significance of the interaction between the corresponding variable is indicated by the elliptical or saddle nature of the contour plots (Muraldihar et al., 2001; Salihu et al., 2011).

Figure 1A represents the interaction between temperature and pressure conditions. Lower and higher levels of both temperature and pressure did not result in higher EOY. Figure 1B shows the 3D plot corresponding to temperature and particle size of the sample, where a moderate interaction between these tested variables occurred but was not significant. In the case of pressure and sample particle size (Figure 1C), the response plot was an elliptical indicating interaction between both with optimum EOY in SC-CO<sub>2</sub> extraction with ethanol as co-solvent.

Furthermore, Figure 2B and 2C show the 3D plots corresponding to temperature and particle size of the sample; and pressure and particle size of the sample, respectively. Lower and higher levels of pressure and sample particle size result in higher FY. Whereas, Figure 2A represents the interaction between temperature and pressure, lower and higher levels of temperature and pressure did not result in higher FY. The shape of the response surface curve showed a moderate interaction between the tested variables. Thus, it can be seen that the optimized combination of selected condition components (temperature, pressure and particle size of the sample) showed the strong synergistic effect on EOY and FY values.

# Conclusion

In this study, SC-CO<sub>2</sub> extraction with ethanol as co-solvent was successful in extracting essential oil and carotenoid (fucoxanthin) from *S. binderi*. An FCCCD under RSM was used to determine the optimal conditions of the three significant factors (temperature, pressure and sample particle size) to yield oil and fucoxanthin extracts. The results demonstrated that optimal EOY and FY by SC-CO2 extraction with ethanol as co-solvent are 10.04 mg/g and 3188.99  $\mu$ g/g achieved at 50°C, 3625 psig and 500  $\mu$ m, respectively which represent the center point (run 13).

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# References

- Abbas, K.A., Mohamed, A., Abdulamir, A.S. and Abas H.A. 2008. A Review on Supercritical Fluid Extraction as New Analytical Method. American Journal of Biochemistry and Biotechnology 4: 345-353.
- Baek, J.K., Kim, S., Lee, G.S. and Shim, J.J., 2004. Density correlation of solubility of CI disperse orange 30 dye in supercritical carbon dioxide. Korean Journal of Chemical Engineering 21(1): 230-235.
- Bari, M.N., Alam, M.Z., Muyibi, S.A. and Jamal, P., 2009. Improvement of production of citric acid from oil palm empty fruit bunches: Optimization of media by statistical experimental designs. Bioresource Technology 100(12): 3113-3120.
- Chen, D., Han, Y. and Gu, Z., 2006. Application of statistical methodology to the optimization of fermentative medium for carotenoids production by *Rhodobacter sphaeroides*. Process Biochemistry 41(8): 1773-1778.
- Dembitsky, V.M. and Maoka, T., 2007. Allenic and cumulenic lipids. Progress in Lipid Research 46(6): 328-375.
- Döker, O., Salgin, U., Yildiz, N., Aydoğmuş, M. and Çalimli, A., 2010. Extraction of sesame seed oil using supercritical CO<sub>2</sub> and mathematical modeling. Journal of Food Engineering 97(3): 360-366.
- Foster, N.R., Singh, H., Yun, S.J., Tomasko, D.L. and Macnaughton, S.J., 1993. Polar and nonpolar cosolvent effects on the solubility of cholesterol in supercritical fluids. Industrial and Engineering Chemistry Research 32(11): 2849-2853.
- Gu, Z., Deming, C., Yongbin, H., Zhigang, C. and Feirong,

G., 2008. Optimization of carotenoids extraction from Rhodobacter sphaeroides. LWT-Food Science and Technology 41(6): 1082-1088.

- Heo, S.J. and Jeon, Y.J., 2009. Protective effect of fucoxanthin isolated from *Sargassum siliquastrum* on UV-B induced cell damage. Journal of Photochemistry and Photobiology B: Biology 95(2): 101-107.
- Hossain, M.B., Barry-Ryan, C., Martin-Diana, A.B. and Brunton, N.P. 2011. Optimisation of accelerated solvent extraction of antioxidant compounds from rosemary (*Rosmarinus officinalis* L.), marjoram (*Origanum majorana* L.) and oregano (*Origanum vulgare* L.) using response surface methodology. Food Chemistry 126(1): 339-346.
- Ismail, H., Fahmi, A., Doolaanea, A.M., Awang, M. and Mohamed, F. 2012. High initial burst release of gentamicin formulated as PLGA microspheres implant for treating orthopaedic infection. International Journal of Pharmacy and Pharmaceutical Sciences 4: 685-91.
- Jaswir, I., Noviendri, D., Salleh, H.M., and Miyashita, K. 2011. Techniques of Extraction and Purification of Fucoxanthin from Brown Seaweeds. In Noorbatcha, I.A., Karim, M.I.A. and Salleh, H.M. (Eds.). Experimental methods in Modern Biotechnology Engineering. 1st ed. Kuala Lumpur: IIUM Press.
- Jaswir, I., Noviendri, D., Salleh, H.M., Taher, M., Miyashita, K. and Ramli, N. 2013. Analysis of fucoxanthin content and purification of all-trans-fucoxanthin from *Turbinaria turbinata* and *Sargassum plagyophyllum* by SiO<sub>2</sub> open column chromatography and reversed phase-HPLC. Journal of Liquid Chromatography and Related Technologies 36(10): 1340-1354.
- Kim, S.M., Jung, Y.J., Kwon, O.N., Cha, K.H., Um, B.H., Chung, D. and Pan, C.H. 2012. A potential commercial source of fucoxanthin extracted from the microalga Phaeodactylum tricornutum. Applied Biochemistry and Biotechnology 166(7):1843-1855.
- Kim, S.M., Shang, Y.F. and Um, B.H., 2011. A preparative method for isolation of fucoxanthin from *Eisenia bicyclis* by centrifugal partition chromatography. Phytochemical Analysis 22(4):322-329.
- Lang, Q. and Wai, C.M. 2001. Supercritical fluid extraction in herbal and natural product studies—a practical review. Talanta 53(4): 771-782.
- Li, P., Xu, L., Mou, Y., Shan, T., Mao, Z., Lu, S., Peng, Y. and Zhou, L. 2012. Medium optimization for exopolysaccharide production in liquid culture of endophytic fungus *Berkleasmium* sp. Dzf12. International Journal of Molecular Sciences 13(9):11411-11426.
- Machmudah, S., Kawahito, Y., Sasaki, M. and Goto, M. 2007. Effect of Supercritical Carbon Dioxide Condition on Extraction of Carotenoids and Seed Oil from Rosehip Fruits. Proceeding of the International Symposium on EcoTopia Science, p. 569-573. Nagoya: EcoTopia Science Institute.
- Maeda, H., Hosokawa, M., Sashima, T., Takahashi, N., Kawada, T. and Miyashita, K. 2006. Fucoxanthin and its metabolite, fucoxanthinol, suppress adipocyte

differentiation in 3T3-L1 cells. International Journal of Molecular Medicine 18(1): 147-152.

- Maeda, H., Tsukui, T., Sashima, T., and Miyashita, K. 2008. Seaweed carotenoid, fucoxanthin, as a multifunctional nutrient. Asia Pacific Journal of Clinical Nutrition 17(S1): 196-199.
- Matsuno, T. 2001. Aquatic animal carotenoids. Fisheries Science 67(5): 771-783.
- Miyashita, K. 2008. Encyclopedia of Cancer. Fucoxanthin, p. 2. New York: Springer-Verlag Berlin Heidelberg
- Mori, K., Ooi, T., Hiraoka, M., Oka, N., Hamada, H., Tamura, M. and Kusumi, T. 2004. Fucoxanthin and its metabolites in edible brown algae cultivated in deep seawater. Marine Drugs 2(2): 63-72.
- Muntari, B., Amid, A., Mel, M., Jami, M.S. and Salleh, H.M. 2012. Recombinant bromelain production in *Escherichia coli*: process optimization in shake flask culture by response surface methodology. AMB Express 2(1): 12
- Muralidhar, R.V., Chirumamila, R.R., Marchant, R. and Nigam, P. 2001. A response surface approach for the comparison of lipase production by *Candida cylindracea* using two different carbon sources. Biochemical Engineering Journal 9(1): 17-23.
- Muthukumaran, P., Gupta, R.B., Sung, H.D., Shim, J.J. and Bae, H.K. 1999. Dye solubility in supercritical carbon dioxide. Effect of hydrogen bonding with cosolvents. Korean Journal of Chemical Engineering 16(1): 111-117.
- Nakazawa, Y., Sashima, T., Hosokawa, M. and Miyashita, K. 2009. Comparative evaluation of growth inhibitory effect of stereoisomers of fucoxanthin in human cancer cell lines. Journal of Functional Foods 1(1): 88-97.
- Naranjo-Modad, S., López-Munguía, A., Vilarem, G., Gaset, A. and Bárzana, E. 2000. Solubility of purified lutein diesters obtained from *Tagetes erecta* in supercritical CO2 and the effect of solvent modifiers. Journal of Agricultural and Food Chemistry 48(11): 5640-5642.
- Nobre, B., Marcelo, F., Passos, R., Beirão, L., Palavra, A., Gouveia, L. and Mendes, R. 2006. Supercritical carbon dioxide extraction of astaxanthin and other carotenoids from the microalga *Haematococcus pluvialis*. European Food Research and Technology 223(6): 787-790.
- Norulaini, N.N., Anuar, O., Abbas, F.M.A., Fatehah, M.O., Omar, A.M., Sahena, F. and Zaidul, I.S.M. 2009. Optimization of supercritical CO<sub>2</sub> extraction of Anastatica hierochuntica. Food and Bioproducts Processing 87(2):152-158.
- Noviendri, D., Jaswir, I., Salleh, H., Taher, M. and Miyashita, K. 2011a. Techniques of Extraction and Purification of Carotenoid (Fucoxanthin) from Brown Seaweed. Workshop on Seaweed Processing for Pharmaceutical Applications. Organized by Bioprocess and Molecular Engineering Research Unit (BPMERU), Department of Biotechnology Engineering, Faculty of Engineering, International Islamic University Malaysia (IIUM), Kuala Lumpur, Malaysia. 16th May 2011, p. 26. Kuala Lumpur: IIUM

- Noviendri, D., Salleh, H.M., Taher, M., Miyashita, K. and Ramli, N. 2011. Fucoxanthin extraction and fatty acid analysis of Sargassum binderi and *S. duplicatum*. Journal of Medicinal Plants Research 5(11): 2405-2412.
- Roh, H.S., Park, J.Y., Park, S.Y. and Chun, B.S. 2006. Isolation of off-flavors and odors from tuna fish oil using supercritical carbon dioxide. Biotechnology and Bioprocess Engineering 11(6): 496-502.
- Roh, M.K., Uddin, M.S. and Chun, B.S. 2008. Extraction of fucoxanthin and polyphenol from *Undaria pinnatifida* using supercritical carbon dioxide with cosolvent. Biotechnology and Bioprocess Engineering 13(6): 724-729.
- Romo-Hualde, A., Yetano-Cunchillos, A.I., González-Ferrero, C., Sáiz-Abajo, M.J. and González-Navarro, C.J. 2012. Supercritical fluid extraction and microencapsulation of bioactive compounds from red pepper (*Capsicum annum* L.) by-products. Food Chemistry 133(3): 1045-1049.
- Salihu, A., Alam, M.Z., AbdulKarim, M.I. and Salleh, H.M. 2011. Optimization of lipase production by *Candida cylindracea* in palm oil mill effluent based medium using statistical experimental design. Journal of Molecular Catalysis B: Enzymatic 69(1): 66-73.
- Santoyo, S., Lloria, R., Jaime, L., Ibanez, E., Senorans, F.J. and Reglero, G. 2006. Supercritical fluid extraction of antioxidant and antimicrobial compounds from *Laurus nobilis* L. Chemical and functional characterization. European Food Research and Technology 222(5-6): 565.
- Simopoulos, A.P. 2002. The importance of the ratio of omega-6/omega-3 essential fatty acids. Biomedicine and Pharmacotherapy 56(8): 365-379.
- Sugawara, T., Matsubara, K., Akagi, R., Mori, M. and Hirata, T. 2006. Antiangiogenic activity of brown algae fucoxanthin and its deacetylated product, fucoxanthinol. Journal of Agricultural and Food Chemistry 54(26): 9805-9810.
- Tanyildizi, M.S., Özer, D. and Elibol, M. 2005. Optimization of α-amylase production by *Bacillus* sp. using response surface methodology. Process Biochemistry 40(7): 2291-2296.
- Turner, C., King, J.W. and Mathiasson, L. 2001. Supercritical fluid extraction and chromatography for fat-soluble vitamin analysis. Journal of Chromatography A 936(1): 215-237.
- Vasapollo, G., Longo, L., Rescio, L. and Ciurlia, L. 2004. Innovative supercritical  $CO_2$  extraction of lycopene from tomato in the presence of vegetable oil as cosolvent. The Journal of Supercritical Fluids 29(1): 87-96.
- Wang, W.J., Wang, G.C., Zhang, M. and Tseng, C.K. 2005. Isolation of fucoxanthin from the rhizoid of *Laminaria japonica* Aresch. Journal of Integrative Plant Biology 47(8): 1009-1015.
- Wu, Y., Cui, S.W., Tang, J. and Gu, X. 2007. Optimization of extraction process of crude polysaccharides from boat-fruited sterculia seeds by response surface methodology. Food Chemistry 105(4): 1599-1605.

- Yan, X., Chuda, Y., Suzuki, M. and Nagata, T. 1999. Fucoxanthin as the major antioxidant in *Hijikia fusiformis*, a common edible seaweed. Bioscience, Biotechnology, and Biochemistry 63(3): 605-607.
- Zhang, P., Xiaohui, L.E.I., Zhongfang, L.E.I., Zhang, Z. and Sugiura, N. 2008. Study on Fucoxanthin Extraction from *Laminaria Japonica* with Ethanol Solution for Health Food Development. Nogyo Shisetsu. Journal of the Society of Agricultural Structures 39(1): 9-16.